Aminolysis of *N*-Tritylamino Acid Benzotriazolyl Esters with Concentrated Aqueous Amines and its Applications in Synthesis

Petros Mamos,^a Elias Dalatsis,^b Costas Athanassopoulos,^b Georgios Balayiannis,^b Dionissios Papaioannou^{b,*} and George W. Francis^c

^aDepartment of Medicine, University of Patras, 265 00 Patras, Greece, ^bDepartment of Chemistry, University of Patras, 265 00 Patras, Greece and ^cDepartment of Chemistry, University of Bergen, Allegaten 41, N-4007 Bergen, Norway

Mamos, P., Dalatsis, E., Athanassopoulos, C., Balayiannis, G., Papaioannou, D. and Francis, G. W., 1998. Aminolysis of *N*-Tritylamino Acid Benzotriazolyl Esters with Concentrated Aqueous Amines and its Applications in Synthesis. - Acta Chem. Scand. 52: 227–231. © Acta Chemica Scandinavica 1998.

Aminolysis of (S)-N-tritylamino acid benzotriazolyl esters with concentrated aqueous amines produces high yields of the corresponding amides which can be readily detritylated with p-toluensulfonic acid or reduced with lithium aluminium hydride to provide chiral ethane-1,2-diamine derivatives.

A variety of biologically important peptides occur in Nature in the form of C-terminal amides. Such peptide amides can be prepared in solution without racemization by direct amidation of the corresponding peptide acids or by amidation of suitably protected C-terminal amino acids followed by assembly of the peptide chain to the required length.1 We have reported that dicyclohexylcarbodiimide (DCC)-mediated activation of N-tritylamino acids with 1-hydroxybenzotriazole (HOBt) produces the corresponding readily isolable esters 1 as an equilibrium mixture of the 'active' ester (1-I) and 'active' amide (1-II) forms.² These esters are highly stable to hydrolysis reactions, are resistant to racemization,³ and show potent acylating abilities and thus constitute valuable intermediates in peptide synthesis.⁴ We now wish to report that esters 1 can be employed in the simple and convenient synthesis of amides using cheap, commercially available, concentrated aqueous (aq.) solutions of simple amines. Others have reported that unrefined preparations of such active esters, N-protected with groups of the urethane type, may be directly converted into primary amides by the action of ammonium hydroxide.⁵

As esters 1 are extremely hydrophobic and exhibit their acylating potential in concentrated solutions, the use of the most concentrated commercially available aq. solutions of amines for amidation is mandatory. Thus, treatment of concentrated solutions of esters 1 in DMF with 30% aq. NH₃, 40% aq. MeNH₂, 33% aq. EtNH₂ or 40% aq. Me₂NH for less than 30 min at 0 °C in most cases produced cleanly (TLC), the corresponding crystal-

line amides 2 in 80–91% yields (Table 1). No sign of hydrolysis of the active esters was detected (TLC) in the amidation reaction mixtures. In accord with earlier observations,² the ester form (1-I) reacts much faster than the amide form (1-II). The latter is rapidly formed in solution when using pure ester form preparations for amidation. In cases where extreme steric hindrance is encountered, e.g., when using Trt-Val-OBt (1b) for amidation, small amounts of the amide form remained unreacted on treatment with both MeNH2 and Me2NH aq. solutions, and were identified as such after isolation by flash column chromatography (FCC) and characterisation by IR and MS (see Experimental). The present method is simple and convenient as it avoids the use of these amines either in their gaseous form or as their corresponding commercially available, but hygroscopic, hydrochlorides.

The trityl (Trt) protective group can be readily removed on brief treatment with $TsOH \cdot H_2O$ in refluxing isopropyl alcohol, a fact demonstrated by the preparation of the tosylates 3a-c, in 75–95% yields. Tosylates like 3 can of course be routinely used for the preparation of analogues of biologically important peptides for structure-activity relationship studies. Furthermore, taking into consideration the compatibility of the Trt group with complex metal hydrides, a property not shared by the protecting groups of the urethane type commonly used for N^{α} -protection of amino acids, amides such as 2 were postulated as convenient sources of chiral ethane-1,2-diamines. In point of fact, treatment of concentrated solutions of, e.g., the amides 2a, b, g and m with LiAlH₄

^{*}To whom correspondence should be addressed.

Fig. 1. Synthetic outline and structures for compounds discussed in the present work.

Table 1. Yields and physical data of α-amino amide and ethane-1,2-diamine derivatives.^a

Compound	Yield (%)	M.p./°C	$[\alpha]_D^{28}$	IR v/cm ⁻¹
Trt-Ala-NHMe (2a)	82	167–168	+32.8	3300, 1664, 1652
Trt-Val-NHMe (2b)	88	190-191	+75.1	3316, 1640
Trt-Val-NMe ₂ (2c)	80	111-112	+ 107.8	3296, 1646, 1634
Trt-Leu-NMe ₂ (2d)	82	105-106	+95.4	3308, 1640, 1628
Trt-Ile-NH ₂ (2e)	87	164-165	+33.4	3452, 3302, 3184, 1682, 1652, 1611
Trt-Ile-NHEt (2f)	84	168-170	+21.1	3332, 1640
Trt-Phe-NHMe (2g)	91	168-169	+21.7	3284, 1682, 1650
Trt-Phe-NMe ₂ (2h)	82	145-146	+ 121.4	3307, 1644, 1632
Trt-Pro-NHMe (2i)	80	134-135	-73.3	3356, 1655, 1642
Trt-His-NH ₂ (2j)	81	121-123	+35.9	3446, 3305, 3234, 1674
Trt-Ser(Bzl)-NMe ₂ (2k)	81	152-153	+ 144.9	3286, 1632
Trt-Glu(Me)-NHEt (21)	84	164-165	+ 56.4	3306, 1732, 1634
Trt-Glu(Me)-NMe ₂ (2m)	85	138-139	+74.3	3292, 1738, 1636, 1626
TsOH · H-Ala-NHMe (3a)	89	145-146	+ 13.7	3328, 1694, 1684 ^b
TsOH · H-Phe-NHMe (3b)	90	192-193	$+33.4^{c}$	3328, 1662
TsOH·H-His-NH ₂ (3c)	75	225-230	-4.5	3428, 1688, 1678
4a	60	180-182	+ 26.3	3295, 3210
4b	65	140-141	-31.0	3415, 3268
4c	74	Oil	+ 49.2	3286
4d	88	Oil	+63.3	3346, 3284

^aAll compounds gave satisfactory microanalytical data (C,H) within ± 0.3 of the calculated values. Yields of reactions are not optimized. Optical rotation values were obtained for 1% solutions in DMF unless otherwise stated. ^bTosylates showed in addition a broad band at 3200–2500 cm⁻¹. ^c1% in MeOH.

in refluxing THF, usually for 1-2 d, produced 60-88% yields of the corresponding diamine derivatives **4a-d**. Worth noting is the reduction of amide **2m** which pro-

duced the diamino alcohol **4d** in less than 1 h, a result which may be attributed to the participation of the newly formed hydroxy function from the reduction of the

Table 2. EI-MS and 200 M Hz ¹H NMR data of α-tritylamino amides.^a

Amide	MS data	NMR data (δ, ppm)
2a	344 (M), 286 (M – CONHMe)	6.538 (1 H, br q, J 5.00 Hz, CONH), 3.281 (1 H, q, J 6.97 Hz, *CH), 2.504 (1 H, br, TrtNH), 2.481 (3 H, d, J 5.00 Hz, NHCH ₃), 1.153 (3 H, d, J 6.97 Hz, CHCH ₃)
2 b	372 (M), 314 (M—CONHMe)	5.877 (1 H, br q, <i>J</i> 4.85 Hz, CONH), 3.092 (1 H, br, *CH), 2.283 (3 H, d, <i>J</i> 4.85 Hz, NHC <i>H</i> ₃), 2.722 (1 H, br, TrtNH) 1.890 (1 H, m, C <i>H</i> Me ₂), 0.942 and 0.932 (6 H, two d, <i>J</i> 6.94 Hz, <i>gem</i> -Me)
2c	343 (<i>M</i> – Me ₂ CH), 314 (<i>M</i> – CONMe)	3.260 (1 H, br d, TrtNH), 3.442 (1 H, m, *CH), 2.373 (3 H, s, NCH ₃), 2.428 (3 H, s, NCH ₃), 2.090 (1 H, m, C H Me ₂), 1.061 (3 H, d, J 6.91 Hz, gem -Me), 0.967 (3 H, d, J 6.91 Hz, gem -Me)
2d	400 (M), 342 (M—CONMe ₂)	3.436 (1 H, br, TrtNH), 3.279 (1 H, br, *CH), 2.474 (3 H, s, NCH ₃), 2.381 (3 H, s, NCH ₃), 2.1–1.9 (3 H, m, CH ₂ and CHMe ₂), 0.942 (6 H, d, J 6.93 Hz, gem -Me)
2e	328 (M – CONH ₂), 86 [EtC(Me)HNH ₂]	3.212 (1 H, br, *CH), 2.661 (1 H, br, TrtNH), 1.611 (1 H, br, CHMe), 1.281 (2 H, m, C H_2 Me), 0.903 (3 H, d, J 6.36 Hz, C H_3 CH), 0.723 (3 H, t, J 7.17 Hz, C H_3 CH $_2$)
2f	328 (M—CONHEt), 258 (TrtNH) 86 [EtCH(Me)NH ₂]	6.138 (1 H, br t, J 5.6 Hz, CONH), 3.173 (1 H, dd, J 3.13 and 4.96 Hz, *CH), 2.852 (2 H, m, NCH ₂), 2.630 (1 H, d, J 5.06 Hz, TrtNH), 1.54–1.06 (3 H, m, MeCH + MeCH ₂), 0.898 (3 H, t, J 7.32 Hz, NCH ₂ CH ₃), 0.880 (3 H,d, J 7.28 Hz, CHCH ₃), 0.746 (3 H, t, J 7.05 Hz, CH ₂ CH ₂)
2 g	362 (<i>M</i> – CONHMe) 343 (<i>M</i> – Ph), 329 (<i>M</i> – PhCH ₂) 120 (PhCH ₂ CHNH ₂)	5.980 (1 H, br q, <i>J</i> 4.98 Hz, CONH), 3.404 (1 H, dt, <i>J</i> 5.29 and 6.04 Hz, *CH), 2.945 (1 H, dd, <i>J</i> 5.29 and 13.5 Hz, PhC <i>H</i> a), 2.726 (1 H, d, <i>J</i> 5.29 Hz, TrtNH), 2.482 (1 H, dd, <i>J</i> 6.04 and 13.5 Hz, PhC <i>H</i> b), 2.351 (3 H, d, <i>J</i> 4.98 and 7.28 Hz, NHC <i>H</i> ₃),
2h	435 (M+H), 362 (M-CONMe ₂) 357 (M-Ph), 343 (M-PhCH ₂) 120 (PhCH ₂ CHNH ₂), 91 (PhCH ₂), 72 (CONMe ₂)	3.618 (1 H, br, TrtNH), 3.429 (1 H, br, *CH), 3.150 (1 H, dd, <i>J</i> 5.53 and 13.03 Hz, PhCHa), 2.870 (1 H, dd, <i>J</i> 8.90 and 12.83 Hz, PhC <i>H</i> b), 2.207 (3 H, s, NCH ₃), 1.758 (3 H, s, NCH ₃)
2 i	70 (M—Trt-MeNCO)	3.922 (1 H, dd, J 1.62 and 7.13 Hz, H[Pro-2]), 3.260 (1 H, m, H[Pro-5a]), 3.030 (1 H, m, H [Pro-5b]), 3.000 (3 H, d, J 4.96, NHC H_3), 1.63–0.95 (4 H, br m, H[Pro-3 and Pro-4])
2 j	445 (M), 443 (M-H ₂) 428 (M-NH ₃), 398 (443-NH ₃ -CO) 372 (M-NH ₂ CHCONH ₂) 130 (M-TrtNHCHCONH ₂)	8.121 (1 H, br s, CONH), 7.52–7.02 (18 H, m, PhH, CONH, ImH), 6.913 (1 H, d, J 2.02 Hz, ImH), 3.546 (1 H, m, *CH), 3.161 (1 H, dd, J 5.82 and 14.36 Hz, ImCHa), 2.911 (1 H, d, J 5.59 Hz, TrtNH), 2.560 (1 H, dd, J 6.12 and 14.36 Hz, ImCHb)
2k	465 (M+H), 392 (M-CONMe ₂) 343 [M-(CH ₂ O+CH ₂ Ph)] 221 (343-HNMe ₂ -Ph), (CONMe ₂)	4.490 (2 H, s, CH_2Ph), 3.976 (1 H, dd, J 8.70 and 5.30 OCHa or OCHb), 3.739 (1 H, m, *CH), 3.563 (1 H, t, J 8.70 OCHa or OCHb), 3.40 (1 H, br, TrtNH), 2.440 and 2.426 (2 × 3 H, s, NMe)

Table 2. (Continued.)

Amide	MS data	NMR data (δ, ppm)
21	431 (M+H), 398 (M-MeOH) 369 (398-Et), 84 (COCHNEt)	5.924 (1 H, br t, <i>J</i> 5.85 Hz, CONH), 3.647 (3 H, s, OMe), 3.223 (1 H, m, *CH), 3.022 (1 H, d, <i>J</i> 5.82 Hz, TrtNH), 2.811 (2 H, dq, <i>J</i> 5.85 and 7.27 Hz, NCH ₂), 2.52–2.15 (2 H, m, CH ₂ CO ₂ Me), 2.08–1.76 (2 H, m, CHCH ₂), 0.879 (3 H, t, <i>J</i> 7.27 Hz, CH ₂ CH ₃),
2m	431 (M+H), 398 (M-MeOH) 358 (M-CONMe ₂), 72 (CONMe ₂)	3.686 (3 H, s, CO_2CH_3), 3.360 (1 H, br, *CH), 2.500 and 2.396 (2 × 3 H, s, NCH_3), 2.90-1.42 (4 H, m, CH_2CH_2)

^aAll MS spectra showed strong peaks at m/z 243 (Trt⁺) and 165 (fluorenyl cation) and weak peaks above 243. However, Trt-Phe-NMe₂ showed the corresponding peaks at m/z 242 (Trt-H) and 166 (fluorenyl cation +H). Only the diagnostically most significant peaks are tabulated. In general, all NMR spectra showed a complex multiplet in the region 7.10–7.40 ppm due to the aromatic protons.

 γ -ester function in the subsequent reduction of the α -amide function. Further work on the preparation of a wider variety of diamines like **4** and their application in asymmetric synthesis is now in progress.

Experimental

General. The benzotriazolyl esters 1 used in the present work were obtained from the corresponding L-amino acids through N-tritylation followed by esterification with HOBt in the presence of DCC.² Capillary melting points were measured on a Büchi SMP-20 apparatus and are uncorrected. Optical rotations were determined with a Carl-Zeiss precision polarimeter. IR spectra were recorded for KBr pellets or neat samples (for oily compounds) on a Perkin-Elmer 16PC FT-IR spectrophotometer. ¹H NMR spectra were obtained at 200.13 MHz on a Bruker AC-200 instrument using CDCl₃ as the solvent and Me₄Si as an internal standard. Mass spectra were obtained on a Fisons VG 7070E mass spectrometer at en electron bombardment energy of 70 eV, using the direct inlet probe at a temperature of 250 °C. FAB mass spectra were recorded on a Fisons-VG ZAB 2f instrument, operated at 8 keV accelerating potential, with an M-SCAN ion gun operated at 10 µA and a 9 keV xenon beam. The matrix was m-nitrobenzyl alcohol with the exception of tosylate 3c for which glycerol was used as the matrix. FCC was performed on Merck silica gel 60 (230–400 mesh) and TLC on Merck silica gel F₂₅₄ films (0.2 mm) precoated on aluminium foil. The solvent systems used were: (A) PhMe-AcOEt (8:2) and (B) CHCl₃-MeOH (9:1). Spots were visualized with UV light at 254 nm and with ninhydrin. THF was distilled from Na-benzophenone.

Amidation procedure. To an ice-cold solution of esters 1 (8 mmol) in DMF (8 ml), 12 mmol each of Et₃N (1.64 ml) and aq. amine were added and the resulting reaction mixture was stirred at 0° C for 15–30 min and then diluted with 5% aq. citric acid. The precipitated

product was extracted twice into EtOAc and the combined organic layers were washed sequentially with H_2O , 5% aq. NaHCO₃, H_2O and brine, and dried (Na₂SO₄). Evaporation of the solvent and addition of Et₂O or Et₂O-hexane (1:1) to the residue caused crystallization of the pure amides **2**. Yields, physical constants and spectral data are tabulated in Tables 1 and 2. In one of the preparations of amide **2c**, the reaction mixture was fractionated by FCC using solvent system A as the eluent, and the unchanged 'active' amide form (R_f 0.54) was isolated and characterised by IR (3332 and 1726 cm⁻¹) and MS (m/z): 475 (M-H), 433 ($M-CHMe_2$), 399 (M-Ph), 314 (M-COBtO) and 243 (Trt).

Detritylation procedure. A suspension of amide 2 (4 mmol) and p-toluenesulfonic acid hydrate (1.1 g, 6 mmol) in 2-propanol (12 ml) was brought to reflux. The resulting solution was kept at reflux temperature for 5 min and then left to attain room temperature. Trituration with Et₂O followed by refrigeration gave the corresponding crystalline tosylates 3 (Table 1). The FAB mass spectra [m/z (% rel. int.)] for tosylates 3 were as follows. 3a: 205 (10.5, [2M+1]), 103 (100, [M+1]), 46 (23.7, $[H_2NCH_2Me+H]$). 3b: 179 (100, [M+1]), 120 (44.7, $[PhCH=CHNH_2+H]$). 3c: 204 (100, [M+1]), 130 (31, $[C_9H_8N]$).

Procedure for LiAlH₄ reduction. To a suspension of LiAlH₄ (0.46 g, 12 mmol) in refluxing THF (4 ml) was added amide 2 (4 mmol) in portions over a period of 30 min and the resulting reaction mixture was further refluxed for 1–2 days, with the exception of amide 2m for which only 1 h was required for completion of the reduction. The reaction mixture was then cooled to 0°C and the excess LiAlH₄ was destroyed by the dropwise addition of a saturated aq. solution of Na₂SO₄. The resulting mixture was filtered and the filtrate evaporated to dryness. The residue was partitioned between EtOAc and brine and the organic phase was dried (Na₂SO₄)

and evaporated to dryness. The residue was subjected to FCC with solvent system B as the eluent to give pure diamines 4 (Table 1). The FAB mass spectra [m/z] (% rel. int.)] for diamines 4 were as follows for 4a: 331 (100 [M+1]), 286 (9, [TrtNH=CHMe]), **4b**: 357 (100, [M+1]). **4c**: 407 (85.7, [M+1]),(100, [TrtNH=CHCH₂Ph]). **4d**: 389 (100, [M+1]), 330 (26.7, [TrtNH=CH(CH₂)₃OH]), 311 (26.7, [TrtNHCH=CHCH=CH₂]).

References

- 1. Somlai, C., Szokan, G. and Penke, B. Synthesis (1995) 683 and references cited therein.
- 2. Barlos, K., Papaioannou, D. and Theodoropoulos, D. Int. J. Peptide Protein Res. 23 (1984) 300.3. Barlos, K., Papaioannou, D. Patrianakou S. and
- Tsegenidis, T. Liebigs Ann. Chem. (1986) 1950.
- 4. Barlos, K., Papaioannou, D. and Sanida, C. Liebigs Ann. Chem. (1984) 1308.
- 5. Chen, S.-T., Wu, S.-H. and Wang, K.-T. Synthesis (1989) 37.
- 6. Mamos, P., Karigiannis, G., Athanassopoulos, C., Bichta, S., Kalpaxis, D., Papaioannou, D. and Sindona, G. Tetrahedron Lett. (1995) 5187 and references cited therein.