

Synthesis of an AZT-HEPT Hybrid and Homologous AzddU Derivatives

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3'-Azido-2',3'-dideoxyuridines **6** and their corresponding α anomers **5** were synthesized by condensation of silylated 6-alkyl and 5,6-dialkyl substituted uracils **2** with methyl 3-azido-5-*O*-*tert*-butyldiphenylsilyl-2,3-dideoxy- α,β -D-erythro-pentofuranoside (**4**). Compounds **5** and **6** were treated with tetrabutylammonium fluoride to obtain the deprotected nucleosides **7** and **8**, respectively.

After the discovery that human immunodeficiency virus (HIV) is the causative agent of acquired immunodeficiency syndrome (AIDS),^{1,2} many 2',3'-dideoxy nucleosides have been synthesized and investigated for their activity against HIV. In particular, 3'-azido-3'-deoxythymidine (AZT)³ is a useful drug against HIV, but also 6-substituted 3'-azido-2',3'-dideoxyuridine (AZddU) derivatives have been of interest as potential drugs.⁴ Recently, acyclic nucleosides of the HEPT type (1-[(2-hydroxyethoxy) methyl]-6-(phenylthio) thymine) have shown high selectivity towards HIV-1,⁵ and it was found that replacement of the sulfur atom with a methylene group resulted in a new series of potent HEPT analogues, of which MKC-442 (6-benzyl-1-ethoxy-methyl-5-isopropyluracil) was found to be extremely potent.⁶

Recently, we have attempted to synthesize a hybrid between HEPT and AZT by condensing 6-phenylthiouracil with an appropriate 3-azido sugar derivative,⁷ but we failed because glycosylation occurred at *N*-3. Now we report that this method instead can be used successfully for the synthesis of a hybrid between AZT and the 6-benzyl analogs of HEPT. In fact, we find that 6-alkyl uracils are generally glycosylated at *N*-1 when condensed with a methyl 3-azido-2-deoxyglycoside.

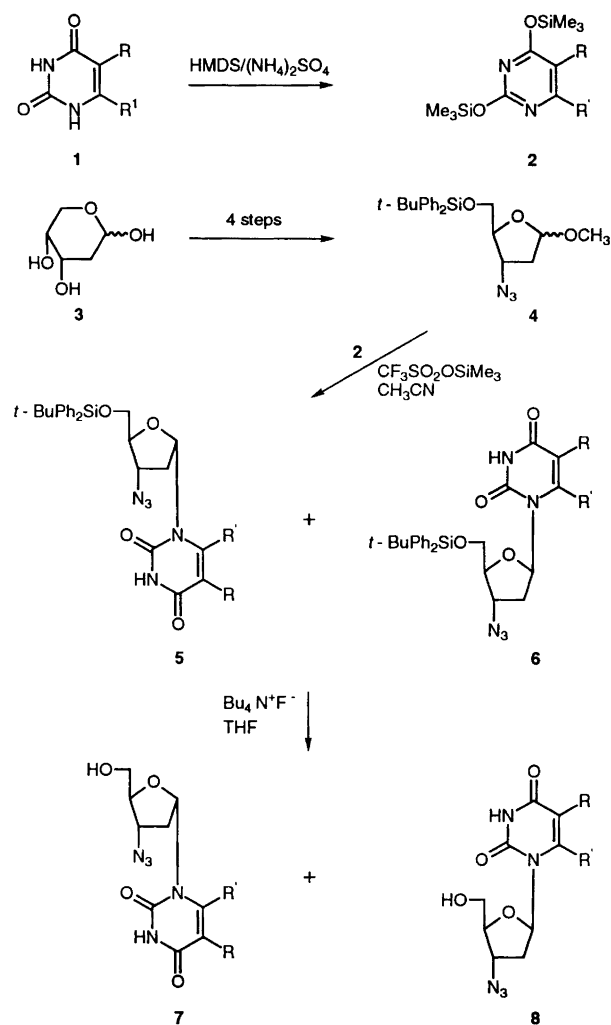
Results and discussion

The starting materials 6-alkyluracils **1c,d**⁸ and 5-ethyl-6-benzyluracil **1e**⁹ were prepared by reaction of ethyl 3-oxovalerate, ethyl butyrylacetate and ethyl 2-ethyl-4-

phenyl-3-oxobutyrate,^{9,10} respectively, with thiourea and sodium ethoxide. The intermediate 2-thiouracils were finally refluxed with chloroacetic acid for 6–18 h. The uracils **1c–e** and the commercially available 6-methyluracil (**1a**) and 5,6-dimethyluracil (**1b**) were silylated¹¹ with 1,1,1,3,3,3-hexamethyldisilazane (HMDS) prior to condensation with methyl 3-azido-5-*O*-*tert*-butyldiphenylsilyl-2,3-dideoxy- α,β -D-erythro-pentofuranoside (**4**). The sugar was prepared in four steps from the commercially available 2-deoxy-D-ribose (**3**) by successive glycosidation¹² with methanolic HCl, selective 5-*O*-silylation with *tert*-butyldiphenylchlorosilane, replacement of 3-hydroxy with an iodo group in a Mitsunobu reaction^{13,14} and finally treatment with sodium azide in dry *N,N*-dimethylformamide.¹⁵ The nucleoside synthesis was accomplished by using trimethylsilyl trifluoromethanesulfonate (TMS triflate) as a Lewis acid catalyst according to the method described by Vorbrüggen *et al.*^{16,17} to give an anomeric mixture of the protected nucleosides **5a,b,e** and **6a,b,e** in 43–64% yield (**5a,6a**: $\alpha/\beta=1:1$; **5b,6b**: $\alpha/\beta=10:7$; **5e,6e**: $\alpha/\beta=7:10$). Subsequent removal of the silyl protecting group with tetrabutylammonium fluoride followed by chromatographic purification afforded the unprotected nucleosides **7a,b,e** and **8,b,e** in 11–29 and 11–21% yields, respectively (Scheme 1).

In the case of the 6-ethyl AZddU derivative, which gave an 3:2 (α/β) anomeric mixture after the coupling reaction, the protected nucleoside was separated into the α - and β -anomers (**5c** and **6c**) by silica gel chromatography in 34 and 25% yield, respectively. Subsequent removal of the silyl protecting group with tetrabutylammonium fluoride followed by chromatographic puri-

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| 1-8 | a | b | c | d | e |
|-----|-----------------|-----------------|-------------------------------|-------------------------------|-------------------------------|
| R | H | CH ₃ | H | H | C ₂ H ₅ |
| R' | CH ₃ | CH ₃ | C ₂ H ₅ | C ₃ H ₇ | CH ₂ Ph |

Scheme 1.

fication afforded the unprotected nucleosides **7c** and **8c** in 23 and 18% yields, respectively. Also in case of the 6-propyl AZddU derivative giving an 4:1 (α/β) anomeric mixture after the coupling reaction, the anomers were separated before deprotection. In all the nucleoside syntheses neither mono- *N*-3 glycosylation nor *N*-1, *N*-3 bisglycosylation was observed when TMS triflate was used as the catalyst in agreement with the literature observation.^{16,18} The new compounds were identified by comparison of similar NMR data,^{9,19} ¹H-COSY and ¹H-nuclear Overhauser effects (NOE) of the compounds **7b,e** and **8b,c,e**. NOE of **7e** confirmed it to be an α anomer. A decisive feature was irradiation of 2'-H at the β face which resulted in 9% NOE in 3'-H and 6% in 1'-H whereas irradiation of 2'-H at the α face gave 3% NOE in 3'-H, 1% in 1'-H and 2% in 4'-H. *N*-1 glycosylation

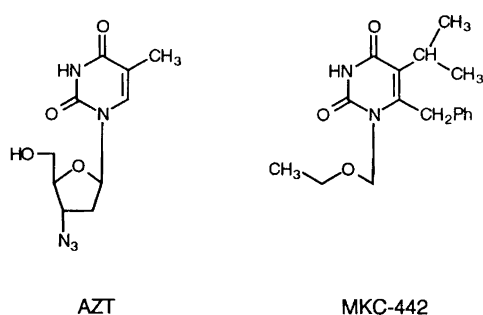


Fig. 1.

was confirmed by 6% NOE in the benzylic CH₂ group when 1'-H was irradiated. NOE of **8e** proved it to be a β anomer as irradiation of 2'-H at the α site resulted in 6% NOE in 1'-H and 2% in 3'-H whereas irradiation of 2'-H at the β site gave 8% NOE in 3'-H and 2% in 1'-H. *N*-1 glycosylation was confirmed by 6% NOE in the benzylic CH₂ group when 1'-H was irradiated.

The nucleosides **7b,e** and **8b** did not show any significant activity in 100 μ M concentration against HIV-1 in MT-4 cells. Compound **8e** showed toxicity against MT-4 cells in 10 μ M concentration, but no activity against HIV-1 was observed at a lower concentration. Expression of HIV in culture medium was quantified by antigen detection ELISA. The same compounds were also devoid of any activity in 100 μ M concentration against herpes simplex virus, type 1 (HSV-1), strain McIntyre when tested in African green monkey kidney cell line Vero.

Experimental

1-(3-Azido-2,3-dideoxy- α -D-erythro-pentofuranosyl)-6-methyluracil (**7a**) and its β -anomer (**8a**).

Typical procedure. 6-Methyluracil (**1a**) (0.78 g, 6.2 mmol) was dissolved in 1,1,1,3,3,3-hexamethyldisilazane (HMDS) (25 ml). (NH₄)₂SO₄ (50 mg, 0.37 mmol) was added and the solution was heated under reflux for 18 h. The solvent was evaporated *in vacuo* to yield the silylated base (**2**) as a colourless oil. Dry MeCN (25 ml) was added to the silylated base, followed by a solution of 3-azidofuranoside **4** (1.70 g, 4.1 mmol) in MeCN (25 ml) and the mixture was cooled to -30°C . Trimethylsilyl trifluoromethanesulfonate (1.1 ml, 6.2 mmol) in MeCN (10 ml) was added dropwise (20 min) to the mixture with stirring. The reaction mixture was stirred at -30°C for 2 h, diluted with CH₂Cl₂ (150 ml), neutralized with cold sat. aq. NaHCO₃ (75 ml), washed with cold sat. aq. NaHCO₃ (2 \times 50 ml) and water (50 ml), dried (Na₂SO₄) and evaporated *in vacuo* to give a crude black product which was subjected to silica gel (150 g) column chromatography with MeOH-CH₂Cl₂ (1:99) to yield 900 mg (43%) of **5a/6a** as a colourless oil ($\alpha/\beta=1:1$). The mixture **5a/6a** (900 mg, 1.8 mmol) was dissolved in THF (30 ml) and 2 ml of 1 M Bu₄NF in THF was added slowly (10 min) at 0°C . The reaction

mixture was stirred for 1 h and the solvent was evaporated *in vacuo*. After silica gel column chromatography with MeOH-CHCl₃ (2:98) afforded the products **7a** as a colourless solid and **8a** as a colourless oil.

Compound 7a. 0.160 g (14%); m.p. 151–153 °C. ¹H NMR (DMSO-d₆/TMS): δ 2.24 (s, 3 H, CH₃), 2.57–2.79 (m, 2 H, 2'-H), 3.32–3.58 (m, 2 H, 5'-H), 4.03 (q, *J* 8.7 Hz, 1 H, 3'-H), 4.21–4.27 (m, 1 H, 4'-H), 4.87 (br s, 1 H, OH), 5.50 (s, 1 H, 5-H), 5.94 (t, *J* 7.3 Hz, 1 H, 1'-H), 11.18 (br s, 1 H, NH). ¹³C NMR (DMSO-d₆/TMS): δ 19.7 (CH₃), 33.6 (C-2'), 59.8 (C-3'), 60.9 (C-5'), 83.4 (C-4'), 85.5 (C-1'), 102.1 (C-5), 150.7 (C-2), 153.2 (C-6), 162.3 (C-4). Anal. C₁₀H₁₃N₅O₄·0.25 H₂O: C, H, N.

Compound 8a. 0.120 g (11%). ¹H NMR (DMSO-d₆/TMS): δ 2.26 (s, 3 H, CH₃), 2.79–2.90 (m, 2 H, 2'-H), 3.57 (t, *J* 5.0 Hz, 2 H, 5'-H), 3.68–3.75 (m, 1 H, 4'-H), 4.32–4.41 (m, 1 H, 3'-H), 4.90 (t, *J* 5.1 Hz, 1 H, OH), 5.50 (s, 1 H, 5-H), 6.01 (dd, *J* 4.4 and 8.8 Hz, 1 H, 1'-H), 11.17 (s, 1 H, NH). ¹³C NMR (DMSO-d₆/TMS): δ 19.5 (CH₃), 35.0 (C-2'), 61.1 (C-3'), 61.3 (C-5'), 84.1 (C-4'), 84.4 (C-1'), 102.2 (C-5), 150.4 (C-2), 153.4 (C-6), 162.2 (C-4). Calc. for C₁₀H₁₃N₅O₄: 267.248. Found 267.250 (MS).

1-(3-Azido-2,3-dideoxy-α-D-erythro-pentofuranosyl)-5,6-dimethyluracil (7b) and its β-anomer (8b). The coupling reaction was performed at –45 °C for 3 h. α/β=10:7 in the crude product. The deprotected nucleosides **7b** and **8b** were purified by silica gel (120 g) column chromatography with Et₂O and obtained as colourless solids.

Compound 7b. 0.244 g (29%); m.p. 166–168 °C. ¹H NMR (DMSO-d₆/TMS): δ 1.81 (s, 3 H, CH₃), 2.24 (s, 3 H, CH₃), 2.49–2.79 (m, 2 H, 2'-H), 3.41–3.57 (m, 2 H, 5'-H), 4.04 (q, *J* 8.7 Hz, 1 H, 3'-H), 4.20–4.27 (m, 1 H, 4'-H), 4.88 (t, *J* 5.6 Hz, 1 H, OH), 6.01 (t, *J* 7.4 Hz, 1 H, 1'-H), 11.17 (br s, 1 H, NH). ¹³C NMR (DMSO-d₆/TMS): δ 10.7 (CH₃), 16.3 (CH₃), 33.6 (C-2'), 59.8, 60.9 (C-3' and C-5'), 83.1, 85.6 (C-1' and C-4'), 107.2 (C-5), 148.0, 150.1 (C-2 and C-6), 163.0 (C-4). Calc. for C₁₁H₁₅N₅O₄: 281.112. Found 281.111 (MS).

Compound 8b. 0.174 g (21%); m.p. 110–112 °C. ¹H NMR (DMSO-d₆/TMS): δ 1.88, (s, 3 H, CH₃), 2.15–2.25 (m, 1 H, 2'-H), 2.26 (s, 3 H, CH₃), 2.79–2.90 (m, 1 H, 2'-H), 3.56–3.60 (m, 2 H, 5'-H), 3.68–3.75 (m, 1 H, 4'-H), 4.34–4.43 (m, 1 H, 3'-H), 4.91 (t, *J* 5.2 Hz, 1 H, OH), 6.09 (dd, *J* 4.5 and 8.7 Hz, 1 H, 1'-H), 11.23 (s, 1 H, NH). ¹³C NMR (DMSO-d₆/TMS): δ 10.8 (CH₃), 16.3 (CH₃), 35.1 (C-2'), 61.1, 61.3 (C-3' and C-5'), 84.0, 84.7 (C-1' and C-4'), 107.4 (C-5), 148.3 (C-2), 150.0 (C-6), 163.0 (C-4). FAB MS (DMSO, 3-nitrobenzylalcohol): *m/z* 282 (*M*+H⁺).

1-(3-Azido-5-O-tert-butylidiphenylsilyl-2,3-dideoxy-α-D-erythro-pentofuranosyl)-6-ethyluracil (5c) and its β-anomer (6c). The reaction was performed at –25 °C for 4 h. α/β=3:2 in the crude product. α and β anomers

were separated by silica gel chromatography with the gradient 0–0.5 % MeOH in CH₂Cl₂.

Compound 5c. 0.860 g (34%). ¹H NMR (DMSO-d₆/TMS): δ 1.00 (s, 9 H, *t*-Bu), 1.12 (t, *J* 7.3 Hz, 3 H, CH₃), 2.48–2.87 (m, 4 H, CH₂ and 2'-H), 3.70–3.84 (m, 2 H, 5'-H), 4.16 (q, *J* 8.8 Hz, 1 H, 3'-H), 4.37–4.43 (m, 1 H, 4'-H), 5.48 (s, 1 H, 5-H), 5.92 (t, *J* 7.2 Hz, 1 H, 1'-H), 7.38–7.65 (m, 10 H, ArH), 11.25 (s, 1 H, NH). ¹³C NMR (DMSO-d₆/TMS): δ 12.2 (CH₃), 18.7 (Me₃C), 25.3 (CH₂), 26.5 (Me₃C), 33.3 (C-2'), 59.7 (C-3'), 63.5 (C-5'), 82.7 (C-4'), 85.1 (C-1'), 100.6 (C-5), 127.7, 127.7, 129.8, 132.7, 135.0 (aryl), 150.8 (C-2), 157.9 (C-6), 162.5 (C-4).

Compound 6c. 0.630 g (25 %). ¹H NMR (DMSO-d₆/TMS): δ 0.99 (s, 9 H, *t*-Bu), 1.12 (t, *J* 7.3 Hz, 3 H, CH₃), 2.25–2.54 (m, 1 H, 2'-H), 2.59 (q, *J* 7.3 Hz, 2 H, CH₂), 2.82–2.92 (m, 1 H, 2'-H), 3.74–3.88 (m, 3 H, 4'-H and 5'-H), 4.37–4.47 (m, 1 H, 3'-H), 5.47 (s, 1 H, 5-H), 6.01 (dd, *J* 3.3 and 8.8 Hz, 1 H, 1'-H), 7.34–7.64 (m, 10 H, ArH), 11.15 (s, 1 H, NH). ¹³C NMR (DMSO-d₆/TMS): δ 12.1 (CH₃), 18.7 (Me₃C), 25.1 (CH₂), 26.4 (Me₃C), 35.2 (C-2'), 61.3 (C-3'), 64.3 (C-5'), 84.0 (C-4'), 84.3 (C-1'), 100.4 (C-5), 127.6, 127.7, 129.3, 129.7, 132.6, 132.8, 134.9, 135.0 (aryl), 150.3 (C-2), 158.1 (C-6), 162.5 (C-4).

1-(3-Azido-2,3-dideoxy-α-D-erythro-pentofuranosyl)-6-ethyluracil (7c). *Typical procedure.* To a solution of **5c** (500 mg, 0.96 mmol) in THF (18 ml) was slowly added (10 min) 1.2 ml of 1 M Bu₄NF in THF at 0 °C. The reaction mixture was stirred for 2 h and the solvent was evaporated *in vacuo*. After silica gel (100 g) column chromatography (gradient from 0–5% MeOH in CHCl₃) afforded the product **7c** as a colourless solid. Yield 0.320 g (23%); m.p. 164–166 °C. ¹H NMR (DMSO-d₆/TMS): δ 1.13 (t, *J* 7.3 Hz, 3 H, CH₃), 2.51–2.84 (m, 4 H, CH₂ and 2'-H), 3.41–3.59 (m, 2 H, 5'-H), 4.01 (q, *J* 8.8 Hz, 1 H, 3'-H), 4.23–4.30 (m, 1 H, 4'-H), 4.85 (t, *J* 5.6 Hz, OH), 5.46 (s, 1 H, 5-H), 5.91 (d, *J* 7.3 Hz, 1 H, 1'-H), 11.19 (s, 1 H, NH). ¹³C NMR (DMSO-d₆/TMS): δ 12.3 (CH₃), 25.3 (CH₂), 33.4 (C-2'), 59.7 (C-3'), 60.9 (C-5'), 83.3 (C-4'), 85.0 (C-1'), 100.3 (C-5), 150.8 (C-2), 158.0 (C-6), 162.5 (C-4). Anal. C₁₁H₁₅N₅O₄: C, H, N.

1-(3-Azido-2,3-dideoxy-β-D-erythro-pentofuranosyl)-6-ethyluracil (8c). Colourless oil. Yield 0.248 g (18%). ¹H NMR (DMSO-d₆/TMS): δ 1.13 (t, *J* 7.3 Hz, 3 H, CH₃), 2.18–2.30 (m, 1 H, 2'-H), 2.59 (q, *J* 7.3 Hz, 2 H, CH₂), 2.79–2.90 (m, 1 H, 2'-H), 3.58 (br s, 2 H, 5'-H), 3.73 (q, *J* 5.9 Hz, 1 H, 4'-H), 4.37 (q, *J* 7.7, 1 H, 3'-H), 4.86 (br s, 1 H, OH), 5.46 (s, 1 H, 5-H), 5.98 (dd, *J* 4.0 and 8.9 Hz, 1 H, 1'-H), 11.19 (br s, 1 H, NH). ¹³C NMR (DMSO-d₆/TMS): δ 12.3 (CH₃), 25.2 (CH₂), 35.2 (C-2'), 61.5, 61.6 (C-3', C-5'), 84.3, 84.4 (C-1', C-4'), 100.4 (C-5), 150.5 (C-2), 158.1 (C-6), 162.5 (C-4). Anal. C₁₁H₁₅N₅O₄·1.0H₂O: C, H, N.

1-(3-Azido-5-O-tert-butyl-diphenylsilyl-2,3-dideoxy- α -D-erythro-pentofuranosyl)-6-propyluracil (5d) and its β -anomer (6d). Compounds **5d** and **6d** were prepared similarly to **5c** and **6c** from 6-propyluracil (**1d**) (1.15, 7.4 mmol). The reaction was performed at -25°C for 4 h. The crude yellow product ($\alpha/\beta=4:1$) was subjected to silica gel (150 g) column chromatography (gradient from 0–0.75% MeOH in CH_2Cl_2) to afford the products **5d** as a colourless and **6d** as a colourless oil.

Compound 5d. 0.717 g (27%). ^1H NMR ($\text{DMSO-d}_6/\text{TMS}$): δ 0.91 (t, J 7.3 Hz, 3 H, CH_3), 1.01 (s, 9 H, *t*-Bu), 1.45–1.67 (m, 2 H, CH_2), 2.36–2.92 (m, 4 H, CH_2 and 2'-H), 3.71–3.85 (m, 2 H, 5'-H), 4.18 (q, J 8.8 Hz, 1 H, 3'-H), 4.38–4.44 (m, 1 H, 4'-H), 5.49 (s, 1 H, 5-H), 5.92 (t, J 7.3 Hz, 1 H, 1'-H), 7.39–7.67 (m, 10 H, ArH), 11.25 (s, 1 H, NH). ^{13}C NMR ($\text{DMSO-d}_6/\text{TMS}$): δ 13.1 (CH_3), 18.7 (Me_3C), 20.5 (CH_2), 26.4 (Me_3C), 33.2 (C-2'), 33.8 (CH_2), 59.5 (C-3'), 63.3 (C-5'), 82.7 (C-4'), 85.2 (C-1'), 101.6 (C-5), 127.7, 127.7, 129.7, 132.6, 132.7, 134.9, 134.9 (aryl), 150.8 (C-2), 156.1 (C-6), 162.3 (C-4).

Compound 6d. 0.163 g (6%). ^1H NMR ($\text{DMSO-d}_6/\text{TMS}$): δ 0.94 (t, J 7.3 Hz, 3 H, CH_3), 1.01 (s, 9 H, *t*-Bu), 1.49–1.65 (m, 2 H, CH_2), 2.27–2.67 (m, 3 H, 2'-H and CH_2), 2.88 (ddd, J 3.2, 3.9 and 5.7 Hz, 1 H, 2'-H), 3.79–3.94 (m, 3 H, 5'-H and 4'-H), 4.43 (q, J 7.6 Hz, 1 H, 3'-H), 6.01 (dd, J 3.3 and 8.8 Hz, 1 H, 1'-H), 7.36–7.67 (m, 10 H, ArH), 11.16 (s, 1 H, NH). ^{13}C NMR ($\text{DMSO-d}_6/\text{TMS}$): δ 13.1 (CH_3), 18.7 (Me_3C), 20.6 (CH_2), 26.4 (Me_3C), 33.7 (CH_2), 35.2 (C-2'), 61.3 (C-3'), 64.4 (C-5'), 84.1 (C-4'), 84.5 (C-1'), 101.4 (C-5), 127.6, 127.7, 129.7, 132.6, 132.8, 134.9, 135.0 (aryl), 150.3 (C-2), 156.3 (C-6), 162.3 (C-4).

1-(3-Azido-2,3-dideoxy- α -D-erythro-pentofuranosyl)-6-propyluracil (7d). Silica gel (125 g) column chromatography (gradient from 0 to 4% MeOH in CHCl_3) afforded a colourless solid. Yield 0.570 g (39%); m.p. 132–134 $^\circ\text{C}$. ^1H NMR ($\text{DMSO-d}_6/\text{TMS}$): δ 0.93 (t, J 7.3 Hz, 3 H, CH_3), 1.53 (hexet, J 7.1 Hz, 2 H, CH_2), 2.46–2.83 (m, 4 H, CH_2 and 2'-H), 3.41–3.60 (m, 2 H, 5'-H), 4.01 (q, J 8.9 Hz, 1 H, 3'-H), 4.24–4.31 (m, 1 H, 4'-H), 4.85 (t, J 5.6 Hz, 1 H, OH), 5.46 (s, 1 H, 5-H), 5.89 (t, J 7.3 Hz, 1 H, 1'-H), 11.19 (s, 1 H, NH). ^{13}C NMR ($\text{DMSO-d}_6/\text{TMS}$): δ 13.2 (CH_3), 20.6 (CH_2), 33.5 (C-2'), 33.8 (CH_2), 59.8 (C-3'), 60.9 (C-5'), 83.4 (C-4'), 85.2 (C-1'), 101.4 (C-5), 150.8 (C-2), 156.3 (C-6), 162.4 (C-4). Calc. for $\text{C}_{12}\text{H}_{17}\text{N}_5\text{O}_4$: 295.300. Found 295.302 (MS).

1-(3-Azido-2,3-dideoxy- β -D-erythro-pentofuranosyl)-6-propyluracil (8d). Silica gel (50 g) column chromatography (gradient from 0 to 2% MeOH in CHCl_3) afforded a colourless oil. Yield 0.128 g (9%). ^1H NMR ($\text{DMSO-d}_6/\text{TMS}$): δ 0.94 (t, J 7.3 Hz, 3 H, CH_3), 1.46–1.61 (hexet, J 7.1 Hz, 2 H, CH_2), 2.19–2.31 (m, 1 H, 2'-H), 2.50–2.57 (m, 2 H, CH_2), 2.83 (ddd, J 4.0, 9.1 and 13.5 Hz, 1 H, 2'-H), 3.57 (t, J 5.5 Hz, 2 H, 5'-H),

3.74 (q, J 6.0 Hz, 1 H, 4'-H), 4.31–4.40 (m, 1 H, 3'-H), 4.87 (t, J 5.6 Hz, 1 H, OH), 5.46 (s, 1 H, 5-H), 5.96 (dd, J 3.9 and 8.8 Hz, 1 H, 1'-H), 11.19 (s, 1 H, NH). ^{13}C NMR ($\text{DMSO-d}_6/\text{TMS}$): δ 13.2 (CH_3), 20.7 (CH_2), 33.7 (CH_2), 35.2 (C-2'), 61.5 (C-3'), 61.5 (C-5'), 84.4 (C-1' and C-4'), 101.3 (C-5), 150.4 (C-2), 156.4 (C-6), 162.3 (C-4).

1-(3-Azido-2,3-dideoxy- α -D-erythro-pentofuranosyl)-6-benzyl-5-ethyluracil (7e) and its β -anomer (8e). The coupling reaction was performed at -45°C for 3 h. $\alpha/\beta=7:10$ in the crude product. The deprotected nucleosides **7e** and **8e** were purified by evaporation of the THF, dissolving in Et_2O , washing with H_2O and drying the organic phase over Na_2SO_4 . Evaporation under reduced pressure and silica gel (100 g) column chromatography with 25% Et_2O in petroleum ether (b.p. 60–80 $^\circ\text{C}$) afforded the nucleosides **7e** and **8e** as colourless oils.

Compound 7e. 0.174 g (11%). ^1H NMR (CDCl_3/TMS): δ 1.04 (t, J 7.3 Hz, 3 H, CH_3), 1.65–1.79 (m, 1 H, 1 H, 2a'-H), 2.36–2.83 (m, 3 H, 2b'-H, CH_2), 3.59 (dd, J 3.4 and 12.5 Hz, 1 H, 5a'-H), 3.79–3.94 (m, 3 H, 5b'-H, 3'-H, CH_2Ph), 4.21 (d, J 21.3 Hz, 1 H, CH_2Ph), 4.29–4.55 (m, 1 H, 4'-H), 5.76 (t, J 7.4 Hz, 1 H, 1'-H), 7.09–7.39 (m, 5 H, ArH). ^{13}C NMR (CDCl_3/TMS): δ 13.7 (CH_3), 19.2 (CH_2), 33.9 (C-2'), 34.5 (CH_2Ph), 59.1 (C-3'), 61.0 (C-5'), 83.9 (C-4'), 86.9 (C-1'), 116.5 (C-5), 127.4, 127.4, 129.2, 134.9 (aryl), 148.4 (C-6), 150.6 (C-2), 163.3 (C-4). FAB MS (CHCl_3 , 3-nitrobenzylalcohol): m/z 372 ($M+H^+$).

Compound 8e. 0.234 g (15%). ^1H NMR (CDCl_3/TMS): δ 1.07 (t, J 7.4 Hz, 3 H, CH_3), 1.34–1.44 (m, 1 H, 2a'-H), 2.36–2.81 (m, 3 H, 2b'-H, CH_2), 3.79–3.94 (m, 4 H, 5'-H, 4'-H, CH_2Ph), 4.27 (d, J 17.2 Hz, 1 H, CH_2Ph), 4.43–4.50 (m, 1 H, 3'-H), 5.76 (t, J 6.6 Hz, 1 H, 1'-H), 7.12–7.41 (m, 5 H, ArH). ^{13}C NMR (CDCl_3/TMS): δ 13.6 (CH_3), 19.2 (CH_2), 34.6 (CH_2Ph), 35.1 (C-2'), 60.5 (C-3'), 62.2 (C-5'), 85.0 (C-4'), 86.7 (C-1'), 117.0 (C-5), 127.3, 127.5, 129.3, 134.7 (aryl), 148.3 (C-5), 150.9 (C-2), 163.1 (C-4). FAB MS (CHCl_3 , 3-nitrobenzylalcohol): m/z 372 ($M+H^+$).

References

- Borre-Sinoussi, F., Chermann, J. C., Rey, F., Nugeyre, M. T., Chamaret, S., Grust, J., Dagnet, C., Axler-Blin, C., Vezinet-Brun, F., Rouzioux, C., Rosenbaum, W. and Montagnier, L. *Science* 220 (1983) 868.
- Gallo, R. C., Salahuddin, S. Z., Popovic, M., Shearer, G. M., Kaplan, M., Haynes, B. F., Palker, T. J., Redfield, R., Oleske, J., Safai, B., White, G., Foster, P. and Markham, P. D. *Science* 224 (1984) 500.
- Mitsuya, H., Weinhold, K. J., Furman, P. A., St. Clair, M. H., Lehrman, S. N., Gallo, R. C., Bolognesi, D., Barry, D. W. and Broder, S. *Proc. Natl. Acad. Sci. USA* 82 (1985) 7096.
- Hřebabecský, H., Holý and De Clercq, E. *Coll. Czech. Chem. Commun.* 55 (1990) 1801.
- Tanaka, H., Baba, M., Hayakawa, H., Sakamaki, T., Miyasaka, T., Ubasawa, M., Takashima, H., Sekiya, K.,

- Nitta, I., Shigeta, S., Walker, R. T., Balzarini, J. and De Clercq, E. *J. Med. Chem.* **34** (1991) 349. Baba, M., De Clercq, E., Tanaka, H., Ubasawa, M., Takashima, H., Sekiya, K., Nitta, I., Umezumi, K., Walker, R. T., Mori, S., Ito, M., Shigeta, S. and Miyasaka, T. *Mol. Pharmacol.* **39** (1991) 805.
6. Yuasa, S., Sadakata, Y., Takashima, H., Sekiya, K., Inouye, N., Ubasawa, M. and Baba, M. *Mol. Pharmacol.* **44** (1993) 895. Baba, M., Shigeta, S., Yuasa, S., Takashima, H., Sekiya, K., Ubasawa, M., Tanaka, H., Miyasaka, T., Walker, R. T. and De Clercq, E. *Antimicrob. Agents Chemother.* **38** (1994) 688. Baba, M., Tanaka, H., Miyasaka, T., Yuasa, S., Ubasawa, M., Walker, R. T. and De Clercq, E. *Nucleosides Nucleotides* **14** (1995) 575.
7. El-Eman, A. A., Pedersen, E. B., Jacobsen, J. P. and Nielsen, C. *Bull. Soc. Chim. Fr.* **130** (1993) 817.
8. Draminski, M. and Fiszer, B. *Roczniki Chem.* **45** (1971) 211.
9. Danel, K., Larsen, E. and Pedersen, E. B. *Synthesis* (1995) 934.
10. Hannick, S. M. and Kishi, Y. *J. Org. Chem.* **48** (1983) 3833.
11. Wittenburg, E. *Z. Chem.* **4** (1964) 303.
12. Motawia, S. M. and Pedersen, E. B. *Liebigs Ann. Chem.* (1990) 599.
13. Kunz, H. and Schmidt, P. *Chem. Ber.* **112** (1979) 3886.
14. Mitsunobu, O. *Synthesis* (1981) 1.
15. Hansen, P. and Pedersen, E. B. *Acta Chem. Scand.* **44** (1990) 522.
16. Vorbrüggen, H., Krolkiewicz, K. and Bennua, B. *Chem. Ber.* **114** (1981) 1234.
17. Vorbrüggen, H. and Höfle, G. *Chem. Ber.* **114** (1981) 1256.
18. Vorbrüggen, H. and Krolkiewicz, K. *Angew. Chem., Int. Ed. Engl.* **14** (1975) 421.
19. Kjærsgaard, U., Pedersen, E. B., Nielsen, C. and El-Torgoman, A. M. *Acta Chem. Scand.* **46** (1992) 1016.

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