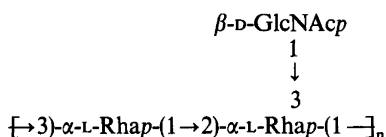


Synthesis of *p*-Nitrophenyl 3-*O*-(2-Acetamido-2-deoxy- β -D-glucopyranosyl)- α -L-Rhamnopyranoside Corresponding to a Fragment of the *Streptococcus* Group A Cell Wall Polysaccharide

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The cell-wall of *Streptococcus* Group A bacteria has been reported¹ to contain the polysaccharide depicted below:



The disaccharide β -D-GlcNAcp-(1 \rightarrow 3)- α -L-Rhap has now been synthesized in the form of the *p*-nitrophenyl glycoside 3, suitable for coupling to proteins. Studies on the antigenic properties of the disaccharide coupled to a protein will be performed.

The free disaccharide has been prepared before² and also, recently, the 8-methoxycarbonyloctyl- α -glycoside.³

The synthesis of 3 is based on silver triflate-assisted glycosidation⁴ of *p*-nitrophenyl 2,4-di-*O*-benzoyl- α -L-rhamnopyranoside⁵ with 3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl bromide.⁴ The protected disaccharide 1, obtained in

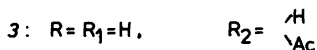
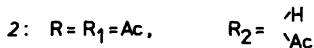
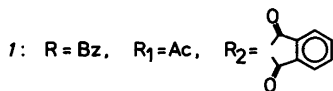
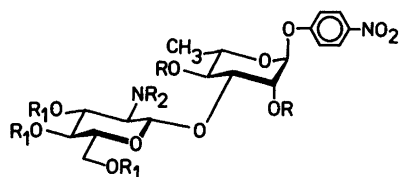
64% yield after chromatography, was subsequently deprotected with hydrazine hydrate⁴ and acetylated to give the crystalline acetate 2 in an 80% yield. De-*O*-acetylation gave the title disaccharide 3 which, after conversion⁶ to the corresponding isothiocyanate derivative, can be used for coupling to proteins.

Experimental. General methods were the same as those reported before.⁷

p-Nitrophenyl 3-*O*-(3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-2,4-di-*O*-benzoyl- α -L-rhamnopyranoside (1). A mixture of *p*-nitrophenyl 2,4-di-*O*-benzoyl- α -L-rhamnopyranoside⁵ (1.20 g, 2.24 mmol), silver triflate (0.82 g, 3.2 mmol) and 2,4,6-trimethylpyridine (0.33 ml, 2.5 mmol) in 1:1 nitromethane-toluene (20 ml) containing powdered 4 Å molecular sieve was stirred and cooled to -20 °C. A solution of 3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl bromide⁴ (1.49 g, 3.0 mmol) in 1:1 nitromethane-toluene (10 ml) was added. After 15 min, more bromide (1.0 g) was added, followed by silver triflate (1.0 g) and 2,4,6-trimethylpyridine (0.4 ml). After 4 h at -20 °C, the mixture was diluted with 1:1 diethyl ether-ethyl acetate and filtered. The filtrate was washed successively with aqueous sodium thiosulfate, water, 2 M aqueous sulfuric acid and aqueous sodium hydrogencarbonate. The residue after drying (MgSO₄) and concentration was purified by column chromatography on silica gel (250 g). Dichloromethane-ethyl acetate-acetone (40:5:1) eluted pure 1 which upon concentration was obtained as a colorless foam (1.35 g, 64%), $[\alpha]_D^{25} -14^\circ$ (c 0.5, CHCl₃). Further elution gave a fraction containing the dehydrohalogenation product of the bromo sugar⁴ (0.82 g).

p-Nitrophenyl 3-*O*-(2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-glucopyranosyl)-2,4-di-*O*-acetyl- α -L-rhamnopyranoside (2). Disaccharide 1 (1.1 g) was treated with a mixture of 95% ethanol and 98% hydrazine hydrate (2:1, 30 ml) at 70 °C for 10 min. After concentration the residue was acetylated (90 °C, 15 min) with pyridine (40 ml) and acetic anhydride (15 ml) and then again concentrated. Purification by column chromatography on silica gel (150 g) using ethyl acetate as eluant gave a solid material which was recrystallized from ethanol. Pure 2 (0.59 g, 80%) was obtained, m.p. 209–213 °C, $[\alpha]_D^{25} -58^\circ$ (c 0.5, CHCl₃). Anal.: C₃₀H₃₈N₂O₁₇: C, H, N.

p-Nitrophenyl 3-*O*-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- α -L-rhamnopyranoside (3). Compound 2 (0.56 g) was treated with 0.1 M methanolic sodium methoxide (20 ml) at room temperature. After 1 h the mixture was neutralized with Dowex 50 (H⁺) and concentrated. The residue, after partitioning between diethyl ether-water and lyophilization of the aqueous phase, was pure 3 (0.36



g, 92 %) as an amorphous solid, $[\alpha]_D -104^\circ$ (c 0.4, H₂O). The 1→3-linkage was demonstrated by methylation analysis – GLC-MS. ¹H NMR (D₂O, 85 °C, external TMS): δ 5.64 (d, $J_{1,2}$ 2.0 Hz, Rha H-1), 4.89 (d, $J_{1,2}$ 7.9 Hz, GlcNAc H-1), 2.03 (acetyl CH₃), 1.20 (d, $J_{5,6}$ 5.4 Hz, Rha CH₃). ¹³C NMR (D₂O, 25 °C, external TMS), δ 176.0 (acetyl C=O), 161.9, 142.9, 126.9, 117.5 (aromatic C), 104.1 (GlcNAc C-1), 98.5 (Rha C-1), 81.1 – 70.7 (ring carbons), 61.6 (GlcNAc C-6), 56.9 (GlcNAc C-2), 23.4 (acetyl CH₃) 18.0 (Rha CH₃).

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