

assisted in obtaining PMR and mass spectra. N. A. was supported in Trondheim by a grant from Hoffmann-La Roche, Basel, to S.L.J.

1. Kelly, M. and Liaaen-Jensen, S. *Acta Chem. Scand.* **21** (1967) 2578.
2. Kelly, M., Norgård, S. and Liaaen-Jensen, S. *Acta Chem. Scand.* **24** (1970) 2169.
3. Ke, B., Imsgard, F., Kjösen, H. and Liaaen-Jensen, S. *Biochim. Biophys. Acta* **210** (1970) 139.
4. Bieman, K., DeJongh, D. C. and Schnoes, H. K. *J. Am. Chem. Soc.* **85** (1963) 1763.
5. Painter, T. J. *Chem. Ind. (London)* **1960** 1214.
6. Hertzberg, S. and Liaaen-Jensen, S. *Phytochemistry* **8** (1969) 1259.
7. Hertzberg, S. and Liaaen-Jensen, S. *Phytochemistry* **8** (1969) 1281.
8. a. Jackman, L. M. and Sternhell, S. *Application of Nuclear Magnetic Resonance Spectra in Organic Chemistry*, 2nd Ed., Pergamon, London 1969; b. Schmidt, K., Francis, G. W. and Liaaen-Jensen, S. *Acta Chem. Scand.* **25** (1971) 2476.
9. Liaaen-Jensen, S. In Isler, O. *Carotenoids*, Birkhäuser, Basel 1971.
10. Arpin, N., Liaaen-Jensen, S. and Trouilloud, M. *Acta Chem. Scand.* **26** (1972) 2524.
11. Gounot, A. M. *Ann. Spéol.* **22** (1967) 23.
12. Liaaen-Jensen, S. and Jensen, A. *Methods Enzymol.* **23** (1971) 586.
13. Kjösen, H., Arpin, N. and Liaaen-Jensen, S. *Acta Chem. Scand. In press.*
14. Norgård, S., Francis, G. W., Jensen, A. and Liaaen-Jensen, S. *Acta Chem. Scand.* **24** (1970) 1460.

Received May 12, 1972.

## Bacterial Carotenoids XL.\* 2'-Hydroxyflexixanthin

MARTHA AGUILAR-MARTINEZ and  
SYNNØVE LIAAEN-JENSEN

*Organic Chemistry Laboratories, Norwegian  
Institute of Technology, University of  
Trondheim, Trondheim, Norway*

The carotenoids of the gliding flexibacteria have been the subject of a number of investigations.<sup>1-6</sup> Flexixanthin (2) and the less abundant deoxyflexixanthin (1) are peculiar to several flexibacteria;<sup>1-3</sup> other flexibacteria produce structurally related carotenoids.<sup>1,2,4-6</sup>

We now report the isolation of flexixanthin (2) and the previously undescribed 2'-hydroxyflexixanthin (3) from a non-gliding bacterium, strain NIVA BR6-64, of uncertain taxonomic position.

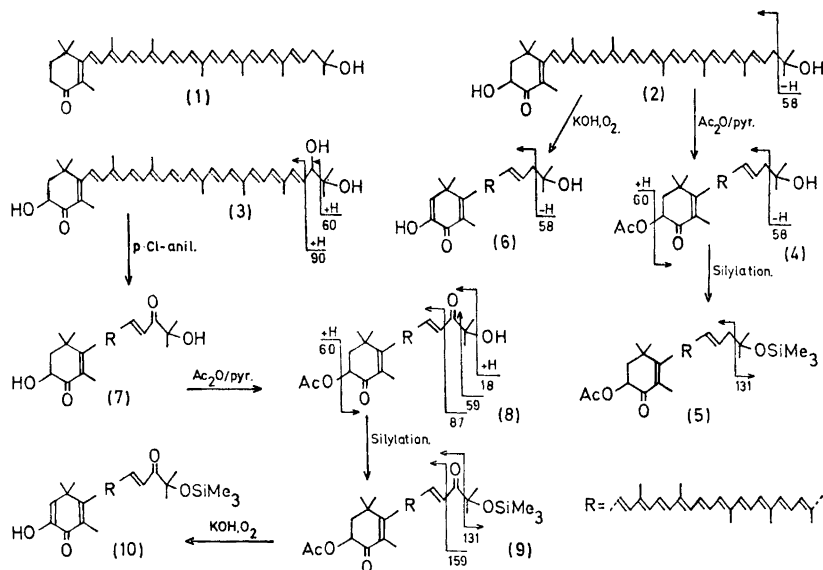
Table 1. Properties of the carotenoids studied.

Carotenoid	$\lambda_{\max}$ in acetone in nm	$R_F$ -values	
		a	b
Deoxyflexixanthin (1)	478	0.52	0.55
Flexixanthin (2)	478,503	0.40	0.27
2'-Hydroxyflexixanthin (3)	478,504	0.19	0.10
Flexixanthin acetate (4)	478	0.50	0.48
4 Trimethylsilyl ether (5)	477	0.68	0.90
Dehydroflexixanthin (6)	478	0.38	0.0
2'-Ketoflexixanthin (7)	500	0.16	0.18
2'-Ketoflexixanthin acetate (8)	499	0.31	
8 Trimethylsilyl ether (9)	500	0.58	0.87
2'-Ketodehydroflexixanthin trimethylsilyl ether (10)	499	0.40	0.0

a. Schleicher & Schüll No. 287 (kieselguhr) paper; 10 % acetone in petroleum ether.

b. Schleicher & Schüll No. 288 (aluminium oxide) paper; 20 % acetone in petroleum ether.

\* No. XXXIX. *Acta Chem. Scand.* **26** (1972) 2526.



Scheme 1.

The carotenoids (14 mg from 110 l of culture, 331 g wet cells) from the acetone extracted cell residue consisted of a minor pigment (0.4 % of the total carotenoids) tentatively identified as deoxyflexixanthin (1), flexixanthin (2, 69 %) and 2'-hydroxyflexixanthin (3, 30 %).

The identification of deoxyflexixanthin (1) was based on its electronic spectrum and chromatographic behaviour (Table 1).

Flexixanthin (2) was identified from electronic and mass spectra; the formation of flexixanthin acetate (4) on acetylation and from the monotrimethylsilyl ether 5 formed by silylation of 4; as well as on the transformation of 2 to dehydroflexixanthin (6) on alkali treatment (Scheme 1, also including important mass-spectrometric fragmentations and Table 1); *cf.* Refs. 3, 7. The derivative 4 could not be chromatographically separated from the corresponding derivative of authentic<sup>3</sup> flexixanthin.

2'-Hydroxyflexixanthin (3) exhibited an electronic spectrum in visible light identical with that of flexixanthin (2). Its mass spectrum showed the molecular ion peak at  $m/e$  598 (consistent with the formula  $C_{40}H_{54}O_4$ ), confirmed by the common losses of 92 and 106 mass units.<sup>8</sup> Diagnostically important peaks were observed at

$M-18(H_2O)$ ,  $M-60$ , and  $M-90$ ; the two latter ions are characteristic<sup>9</sup> of the acyclic end of plectanixanthin (3',4'-didehydro-1',2'-dihydro- $\beta,\gamma$ -caroten-1',2'-diol<sup>10</sup>). The chromatographic behaviour (Table 1), electronic and mass spectra can all be accommodated by structure 3. Supporting evidence was obtained from the chemical transformations summarized in Scheme 1. Oxidation of 3 with *p*-chloranil<sup>11</sup> gave a single product 7. The bathochromic shift<sup>12</sup> and decrease in polarity relative to 3 supported the presence of an allylic hydroxy group in 2'-position in 3. Acetylation of the diketone 7 gave the acetate 8 with higher  $R_F$  value and unchanged electronic spectrum. The mass spectrum of 8 had the molecular ion at  $m/e$  638, confirming the formation of a monoacetate and thus demonstrating the presence of a second secondary or primary hydroxy group in 3. In addition to the common  $M-92$  and  $M-106$  ions, fragment ions of 8 were noted at  $M-60$  (acetic acid) and the expected  $M-59$  and  $M-87$  ions associated with cleavages of the bonds  $\alpha$  to the 2'-keto group. Prominent  $M-16$ <sup>9</sup> and  $M-18$  ions were also observed. The diketo-monoacetate (8) provided a trimethylsilyl ether 9 on silylation<sup>7</sup> with the expected polarity (Table 1).

The mass spectrum of 9 showed the molecular ion at  $m/e$  710 (consistent with the formula  $C_{45}H_{62}O_5Si$ ) and diagnostically useful peaks at  $M-60$  (acetic acid),  $M-131$  ( $C_6H_{16}OSi$ ) and  $m/e$  131. Finally the  $\alpha$ -ketol arrangement of 3 was demonstrated by weak alkali treatment of 9 on the micro scale leading to the acidic diosphenol 10, completely retained on alumina paper; hydrolysis of the tertiary silyl ether group is expected to require stronger conditions.<sup>7</sup> By the new nomenclature<sup>10</sup> 2'-hydroxy-flexixanthin is 3,1',2'-trihydroxy-3',4'-dihydro-1',2'-dihydro- $\beta,\psi$ -caroten-4-one.

The carotenoid analysis may indicate a relationship between the present non-gliding bacterium and the gliding flexibacteria. Whereas nutritional and physiological studies by Ormerod and Kristensen<sup>13</sup> have not provided information facilitating further classification, the % GC ratio falls within the range characteristic of *Flexibacteriales*.<sup>13,14</sup> However, although flexixanthin (2) has not yet been encountered outside flexibacteria it should be mentioned that carotenoids structurally related to 2 and 3 are reported from *Mycobacterium phlei* strain Vera<sup>12</sup> (the aglycone of 4-ketophleixanthophyll), from species of *Myxobacteriales*: *Sorangium compostitum* (the aglycone of myxobacton ester)<sup>15</sup> and *Stigmatella aurantiaca*.<sup>16</sup> These carotenoids differ from 2 and 3 by lacking the  $\alpha$ -ketol arrangement of the  $\beta$ -ring.

*Experimental.* Strain NIVA BR6-64 from the collection of the Norwegian Institute of Water Research was obtained from Docent J. G. Ormerod, Botanical Institute, University of Oslo. The cultivation<sup>17</sup> was carried out in a pilot plant fermentor in a trypton-sucrose medium.

Methods and instruments commonly used in this laboratory were employed.<sup>18,19</sup> The carotenoids were extracted with acetone, saponification being avoided in the isolation procedure, and separated by chromatography on preparative silica gel G plates using 30 % acetone in petroleum ether.

*Acknowledgement.* We thank Docent J. G. Ormerod for providing the culture and suggesting the project, and Docent K. E. Eim-

hjellen, Department of Biochemistry, this Institute, for arranging the cultivation. M.A.-M. was supported by a grant from the Norwegian Agency for International Development.

1. Fox, D. L. and Lewin, R. A. *Can. J. Microbiol.* **9** (1963) 753.
2. Aasen, A. J. and Liaaen-Jensen, S. *Acta Chem. Scand.* **20** (1966) 811.
3. Aasen, A. J. and Liaaen-Jensen, S. *Acta Chem. Scand.* **20** (1966) 1870.
4. Aasen, A. J. and Liaaen-Jensen, S. *Acta Chem. Scand.* **20** (1966) 2322.
5. Aasen, A. J., Liaaen-Jensen, S. and Borch, G. *Acta Chem. Scand.* **26** (1972) 404.
6. Aasen, A. J. *Thesis*, Royal Institute of Technology, Stockholm 1972.
7. McCormick, A. and Liaaen-Jensen, S. *Acta Chem. Scand.* **20** (1966) 1989.
8. Vetter, W., Englert, G., Rigassi, N. and Schwieter, U. In Isler, O. *Carotenoids*, Birkhäuser, Basel 1971, Chapter IV.
9. Enzell, C. R., Francis, G. W. and Liaaen-Jensen, S. *Acta Chem. Scand.* **23** (1969) 727.
10. IUPAC Tentative rules for the Nomenclature of Carotenoids, *Biochem. In press*.
11. Liaaen-Jensen, S. *Acta Chem. Scand.* **19** (1965) 1166.
12. Hertzberg, S. and Liaaen-Jensen, S. *Acta Chem. Scand.* **21** (1967) 15.
13. Ormerod, J. G. and Kristensen, T. *Unpublished results*.
14. Mandel, M. and Lewin, R. A. *J. Gen. Microbiol.* **58** (1969) 171.
15. Kleinig, H., Reichenbach, H., Achenbach, H. and Stadler, J. *Arch. Mikrobiol.* **78** (1971) 224.
16. Kleinig, H. and Reichenbach, H. *Arch. Mikrobiol.* **68** (1969) 210.
17. Hanto, B., Larsen, M. and Sem, R. Student report, Department of Biochemistry, Norwegian Institute of Technology 1971.
18. Liaaen-Jensen, S. and Jensen, A. *Methods Enzymol.* **23** (1971) 586.
19. Kjosen, H., Arpin, N. and Liaaen-Jensen, S. *Acta Chem. Scand. In press*.

Received May 20, 1972.