by treatment with excess of 0.1 N NaOH and
back-titration by 0.1 N HCl. Hydrolysis of
the crude polysaccharide mixture (2 N H₂SO₄,
18 h, 100°) gave the monosaccharides of
Table I; identified by chromatography in
(A), (B), (C), and by measuring [α]D after
separation of thick paper.

Degradation with pectinase. Crude poly-
saccharide was hydrolysed (1 N H₂SO₄, 6 h,
100°), neutralised with BaCO₃, and separated
on De-Aedite (formate form) in a neutral
and an acid fraction. The latter was incubated
for 18 h at 37° with pectinase (pure, Sigma).
The enzyme was removed by heating a few
minutes at 100° and filtered through Millipore
Membrane filter. Galacturonic acid and small
amounts of galactose and arabinose were
liberated. Incubation of crude polysaccharide
with pectinase under the same conditions
as above gave galacturonic acid, galactose,
glucose, and arabinose as main components.

Fractionation of the crude polysaccharide
was carried out on DEAE cellulose (Whatman
DE 11). The polysaccharide (0.1–0.2 g) was
dissolved in water and poured in a column
(2 x 30 cm) in phosphate form.8 The column
was eluted as indicated in Fig. 1. The composi-
tion of the various fractions is given in Table I.

In 1957, we reported the presence of 3-
carbomethoxypropylglucosinolate** (I)

\[
\text{MeO}\overbrace{\text{O}}^{\text{N}}\text{C}_8\text{H}_7\text{O}_5\text{S}^\ominus\text{OSO}_3^\ominus
\]

(1)

in seeds of the cruciferous species Erysimum
rupestre DC., E. ochroleucum DC., and E.
pumilum DC. The seed materials employed
were propagated in the Botanic Garden of
the University of Copenhagen from small
seed specimens donated and labelled by a
German university botanic garden. Un-
fortunately, the seed-producing plants
were not subjected to botanical control at
the time of the cultivation.

In connection with other studies, the
three seed samples were sown again in 1966,
and the resulting plants were now kindly
investigated botanically by Professor C.
Favarger, Institut de Botanique, Université
de Neuchatel, who established the
three originally studied "species" as identi-
cal and different from authentic specimens of
E. rupestre DC., E. ochroleucum DC, and
E. pumilum DC. According to Professor
Favarger's diagnosis, the original studies
were carried out on Erysimum odoratum
Ehrh. which should therefore be regarded

* On leave of absence from Freie Universität
Berlin.

** The trivial name "glucoerypsin" was
then proposed for this thioglucoside. Subse-
quent introduction, however, of a semisys-

tematic "glucosinolate"-nomenclature has ren-
ered the original and ambiguous nomenclature
superfluous and undesirable (cf. Ref. 2).
as the first, and so far only source of the glucosinolate (I).*

The present case may serve as a warning. Whenever sets of chemical and biological parameters (such as a botanical name) are linked together, care and caution must be exercised, also in the biological identification, in order to make the combined information meaningful.

Through the good offices of Professor Favarger, a small amount of seed of the authentic *Erysimum ochroleucum* DC. was put at our disposal and propagated in the Botanic Garden of the University of Copenhagen. Chromatographic analysis of the seed revealed its contents of two glucosinolates, identified as 3-methylthio-propyl; and the corresponding 3-methyl-sulphinylpropyl-glucosinolate, the latter with (R)-configuration around the sulphone dioxide grouping. The identification was effected by enzymic hydrolysis to the corresponding thiocyanates and conversion of the latter into crystalline thiourea derivatives which were compared with authentic specimens (see Experimental).

In conclusion, seeds of *Erysimum odoratum* Ehrh. contain (I) as the predominant glucosinolate. *E. ochroleucum* DC. does not contain (I) but possesses a distinctly different pattern as described above.

Experimental. Seeds of *Erysimum ochroleucum* DC. (25 g) were ground, defatted, and extracted with 70 % methanol. The extract residue was dissolved in water (500 ml) and passed through a column containing anionotropic alumina (Woelm, 100 g). The column was rinsed with water, and the glucosinolates were eluted with 1 % NaOH-solution; 25-ml fractions were collected. Fractions Nos. 5–8 were combined, diluted with an equal volume of citrate buffer (pH 6.8), and subjected to enzymic hydrolysis by the addition of a crude myrosinase solution and a trace of ascorbic acid. After standing for several hours at room temperature, the reaction mixture was extracted three times with ether.

The dried ether extract was mixed with a methanolic solution of ammonia. After an hour, the solution was taken to dryness, and the crystalline residue was recrystallized, first from ethyl acetate:petroleum ether, and then from water. The colourless crystals had m.p. 67–68° (uncorr.) alone, or in admixture with an authentic specimen of 1-(3-methylthiopropyl)thiourea.\(^2\) The naturally derived and the synthetic thiourea exhibited coinciding infra-red spectra.

The aqueous phase from the ether extraction was separated with three small portions of chloroform. To the dried solution was added a slight excess of aniline, the solution was set aside for 2 h at room temperature, and then concentrated to dryness. The crystalline residue was recrystallized, first from ether:petroleum ether, then from water to give colourless crystals, m.p. 134° (uncorr.), alone or mixed with an authentic specimen of (R)-1-(3-methylsulphinylpropyl-3-phenyl-thiourea.\(^4\)

The two specimens exhibited identical IR-spectra. The rotation, \([\alpha]_D^{20} = -45° (c 1.5, EtOH), was numerically slightly lower than that previously reported (-54° ± 2)\(^5\).

The kind assistance of the Botanic Garden of the University of Copenhagen in the production of seed material for the present investigation is gratefully acknowledged. The authors are much indebted to Professor C. Favarger for his expert advice on the botanical identity of the various *Erysimum* species and for a generous seed sample of authentic *E. ochroleucum* DC. The support of the present investigation by *Statens Teknisk-Videnskabelige Fond* is gratefully acknowledged.


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*Herbarium vouchers of plants grown from seed employed in the chemical work have been deposited in the Botanic Museum of the University of Copenhagen.*


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