

Chlorination of Cholesterol in Aqueous Solution: Isolation of a *trans*-Diequatorial Chlorohydrin

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In connection with a study of the reactions which wood extractives undergo during the bleaching of wood pulp, the chlorination of cholesterol in aqueous *t*-butanol solution was examined.¹ Besides the wellknown reaction products, 5 α ,6 β -dichloro-cholestan-3 β -ol,² 5 α -chloro-cholestan-3 β ,6 β -diol and 6 β -chloro-cholestan-3 β ,5 α -diol,³ we obtained two ketonic oxidation products, 5 α -chloro-cholestan-3 β -ol-6-one and 6 β -chloro-cholestan-5 α -ol-3-one, as well as an unidentified compound with the formula C₂₇H₄₄Cl₂O and a new chlorohydrin. The last compound was isolated as its acetate (*A*) in a yield of 15%. The acetate *A* was also obtained by chlorination of cholesterol acetate. This communication describes its identification.

The compound *A* analysed for C₂₉H₄₈ClO₂. Its IR-spectrum showed the presence of acetate and hydroxyl groups. The mass spectrum indicated that it was a 5,6-substituted chlorohydrin as it showed great similarities with the spectra of other 5,6-substituted chlorohydrins. Alkaline saponification yielded 5 β ,6 β -epoxy-cholestan-3 β -ol. The hydroxyl group of the chlorohydrin was therefore in either the 5 β or the 6 β -position. As the compound did not react during attempted acetylation with acetic anhydride and pyridine the hydroxyl group was evidently tertiary and therefore linked to the 5 β -position. Compound *A* is thus IIa or the isomer with a 6 β -chloro group instead of the 6 α -chloro group.

The NMR frequencies of *A* are shown in Table 1, which for comparison also includes those of 3 β -acetoxy-5 α -chloro-cholestan-6 β -ol (IIIa). The half-band widths of the NMR signals are especially informative. The width is considerably larger for an axial proton than for the corresponding equatorial one.⁴ Consequently, the signal width for the axial 3 α -proton in IIIa (25 cps) is considerably larger than the width of the signal of compound *A* which is attributed

Table 1. The proton magnetic signals^a from 3 β -acetoxy-6 α -chlorocholestan-5 β -ol (IIa).

Shift (δ) ppm		Character ^b	Assignment
CDCl ₃ ^c	C ₆ H ₆		
0.66	(0.68)	0.55 s	18-CH ₃
0.88	(0.87)	0.97 d, $J_c = 5.5$	26,27-CH ₃
1.00	(1.30)	0.97 s	19-CH ₃
2.09	(2.03)	1.58 s	3 β -CH ₃ COO
2.98		5.95 broad	5 α -OH
4.4	(4.0 ^d)	4.4 m, $W = 20$	6 α -H
5.35	(5.4 ^e)	5.3 m, $W = 8$	3 α -H

^a 60 Mc.

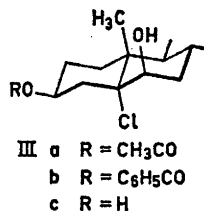
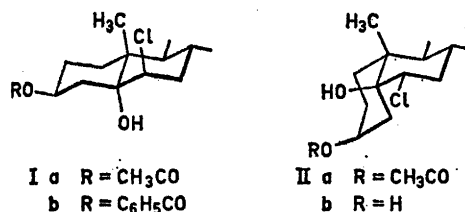
^b s is singlet; d, doublet; m, multiplet. J_c is the coupling constant and W , the half-band width, both in cps.

^c The figures in brackets refer to the signals from 3 β -acetoxy-5 α -chloro-cholestan-6 β -ol (IIIa). If not otherwise stated, the signal characters and assignments are the same as those for (IIa).

^d $W = 6$ cps. Assigned to the equatorial 6 α -H.

^e $W = 25$ cps. Assigned to 3 α -H, which in this compound is axial.

to the equatorial 3 α -proton (8 cps). The *A* proton which is geminal to the chloro atom gives a signal whose width is 20 cps. If compared with 6 cps for the equatorial 6 α -proton of IIIa it is evident that the proton in *A* is axial, which agrees with a 6 α -chloro group.



The compound *A* is then 3 β -acetoxy-6 α -chloro-cholestan-5 β -ol (IIa) in which the 5,6-substituents are *trans*-diequatorial.

Chlorination of cholesterol thus yields 6 α -chloro-cholestan-3 β ,5 β -diol (IIb). The possibility that it is formed by rearrangement⁵ of 5 α -chloro-cholestan-3 β ,6 β -diol (IIIc) was excluded by the finding that IIIc is stable under the reaction conditions.

Previously, Alt and Barton⁶ had found that the chlorination and bromination of Δ^2 - and Δ^5 -cholestene formed small amounts of *trans*-diequatorial dihalo products together with the *trans*-diaxial compounds.

The 5 β -hydroxyl and the 3 β -acetate groups in IIa are suitably situated for the formation of a hydrogen bridge. Such an intermolecular bond explains why the stretching frequency for the acetate carbonyl is unusually low, 1707 cm⁻¹ (in KBr), compared with that for other steroidal 3-acetates, 1719–1728 cm⁻¹ (in CHCl₃).⁷ It is well known that hydrogen bonding decreases the carbonyl frequency.⁸ The hydrogen bond and the 5 β -structure explain why IIa moved considerably faster on thin layer chromatography than the chlorohydrins Ia and IIIa and also cholesteryl acetate: its hydroxyl is masked by the hydrogen bridge and its acetate group is in the less exposed axial position.

The optical rotation of 3 β -benzoxy-5 α -chloro-cholestan-6 β -ol (IIIb) was found to be $[\alpha]_{578} - 21^\circ$ (c, 0.1; CHCl₃) which differs considerably from that reported earlier, $[\alpha]_{Na} \pm 0$.⁹

Experimental. 3 β -Acetoxy-6 α -chloro-cholestan-5 β -ol (IIa) was obtained from chlorination of cholesterol or its acetate as described in Ref. 1. Recrystallisations from ethanol yielded a chlorine-containing product with m.p. 127–129°C and $[\alpha]_{578} + 39^\circ$, $[\alpha]_{365} + 110^\circ$ (c, 2.0; CHCl₃). (Found: C 72.6; H 10.3. Calc. for C₂₉H₄₉ClO₂: C 72.4; H 10.3). The proton magnetic signals are shown in Table 1. The mass spectrum gave the molecular peaks at *m/e* 480 and *m/e* 482, and peaks due to loss of HCl, H₂O, CH₃COOH, and combinations thereof. The spectra of Ia, IIa, Ib, and IIIb were identical below *m/e* 366 (the molecular ions minus HCl, H₂O, and AcOH). The IR

spectrum showed peaks at 3430, 1707, 1265, and 1035 cm⁻¹ (KBr).

The compound IIa (104 mg) was dissolved in a methanol solution of potassium hydroxide (0.5 N, 20 ml). The solution was left for 17 h at room temperature. Water was added and the crystals obtained (59 mg) were collected and recrystallised (methanol). The product, m.p. 132–133°C, $[\alpha]_{578} + 10^\circ$ was indistinguishable from 5 β ,6 β -epoxy-cholestan-3 β -ol (mixed m.p., IR, and NMR).

A solution of 5 α -chloro-cholestan-3 β ,6 β -diol (IIIc) (0.2 g) in 0.2 M hydrochloric acid/t-butanol (20 ml/80 ml) was kept at room temperature. Thin layer chromatography, which was carried out under such conditions that IIb and IIIc should be separated, showed that IIIc did not react in 7 days.

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1. Lindgren, B. O. *Svensk Papperstid. In print.*
2. Barton, D. H. R. and Miller, E. *J. Am. Chem. Soc.* **72** (1950) 370.
3. Ueno, Y. *J. Pharm. Soc. Japan* **72** (1952) 1620; [*Chem. Abstr.* **47** (1953) 8764]; Mori, S. *J. Chem. Soc. Japan* **64** (1943) 981; **71** (1950) 600; [*Chem. Abstr.* **41** (1947) 3807; **45** (1951) 9069].
4. Bhacca, N. S. and Williams, D. H. *Applications of NMR Spectroscopy in Organic Chemistry, Illustrations from the Steroid Field*, Holden-Day 1964, p. 79.
5. Wendler, N. L. In Mayo, P., de, *Molecular Rearrangement*, Interscience, New York 1964, p. 1130.
6. Alt, G. H. and Barton, D. H. R. *J. Chem. Soc.* **1954** 4284.
7. Jones, R. N. and Herling, F. *J. Org. Chem.* **19** (1954) 1252.
8. Rao, C. N. R. *Chemical Applications of Infrared Spectroscopy*, Academic, New York 1963, p. 209.
9. Spring, F. S. and Swain, G. *J. Chem. Soc.* **1939** 1356.

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