

## The Action of Trypsin on Casein

JOHN GRAAE and  
E. SKIPPER RASMUSSEN*Universitetets Plantefysiologiske Laboratorium, Copenhagen, Denmark*

The proteolytic enzymes from pancreas have been studied extensively ever since their discovery and several reaction mechanisms have been proposed to explain the observed kinetic data. Recently the mechanism of action of both trypsin and chymotrypsin have been shown to be more complex than was previously suggested<sup>1,2</sup>. Further data related to the mechanism of trypsin action are presented here.

Crystalline trypsin was kindly placed at our disposal by *Novo Terapeutisk Laboratorium*, Copenhagen, Denmark.

The substrate was a commercial »Casein nach Hammarsten». A stock solution was prepared according to Kunitz<sup>3</sup>, except that instead of buffer the medium was 0.2 M KCl and KOH was added to adjust pH. This solution was diluted with 0.2 M KCl prior to experiment in order to give the desired concentration.

*Table 1.* The table gives the constants A and B. C is titration value after complete hydrolysis and can be taken as a measure of initial substrate concentration. Enzyme concentration equals 6.5 mg/l in all experiments. (Calculations based on log<sub>10</sub>)

| -A min | B min | C     |
|--------|-------|-------|
| 29.2   | 140.0 | 1 080 |
| 16.4   | 83.4  | 620   |
| 8.0    | 44.4  | 340   |
| 4.75   | 26.1  | 180   |

The reaction was followed by continuous titration as previously described<sup>2</sup>. The pH was kept constant by the addition of KOH in minute quanta from a syringe; 1 titration unit equalled 0.0294  $\mu$ equiv. The reaction mixture consisted of 30 ml casein solution and the reaction was initiated by the addition of 1 ml trypsin solution (20.1 mg/100 ml). Experiments were performed at pH = 7.00 and temperature = 25.3°C.

The reaction can be described by a combination of a zero order term and a first order term:

$$t = A \cdot a + B(-\log(1-a)),$$

*Table 2.* The table gives measured and calculated times,  $t_m$  and  $t_c$ , and degree of reaction,  $a$ .  $a$  is determined as the ratio between titration value at time  $t$  and titration value after complete hydrolysis. The calculations are based on the constants given in Table 1.

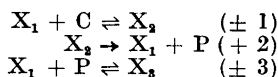
| $t_c$ | $t_m$ | $a$    | $t_c$ | $t_m$ | $a$    | $t_c$ | $t_m$ | $a$    | $t_c$ | $t_m$ | $a$    |
|-------|-------|--------|-------|-------|--------|-------|-------|--------|-------|-------|--------|
| 2.0   | 2     | 0.0593 | 1.7   | 2     | 0.0774 | 0.6   | 1     | 0.0471 | 0.8   | 1     | 0.1111 |
| 4.3   | 4     | 0.1213 | 3.7   | 4     | 0.1613 | 1.8   | 2     | 0.1412 | 1.8   | 2     | 0.2222 |
| 6.2   | 6     | 0.1657 | 6.1   | 6     | 0.2419 | 2.7   | 3     | 0.2000 | 2.8   | 3     | 0.3166 |
| 8.0   | 8     | 0.2065 | 8.5   | 8     | 0.3129 | 3.9   | 4     | 0.2706 | 4.0   | 4     | 0.4055 |
| 9.6   | 10    | 0.2389 | 9.9   | 10    | 0.3516 | 5.1   | 5     | 0.3324 | 5.1   | 5     | 0.4777 |
| 12.0  | 12    | 0.2843 | 12.2  | 12    | 0.4065 | 5.9   | 6     | 0.3676 | 6.0   | 6     | 0.5277 |
| 14.0  | 14    | 0.3175 | 14.4  | 14    | 0.4516 | 7.2   | 7     | 0.4205 | 7.4   | 7     | 0.5944 |
| 15.7  | 16    | 0.3453 | 16.6  | 16    | 0.4952 | 8.3   | 8     | 0.4617 | 8.4   | 8     | 0.6333 |
| 17.1  | 18    | 0.3667 | 18.3  | 18    | 0.5242 | 10.7  | 10    | 0.5412 | 9.3   | 9     | 0.6666 |
| 25.2  | 25    | 0.4731 | 20.7  | 20    | 0.5629 | 12.6  | 12    | 0.5941 | 10.0  | 10    | 0.6888 |
| 31.3  | 32    | 0.5380 | 25.7  | 25    | 0.6306 | 15.8  | 15    | 0.6647 | 12.6  | 12    | 0.7611 |
| 42.0  | 44    | 0.6296 | 30.7  | 30    | 0.6855 | 20.5  | 18    | 0.7471 | 14.2  | 14    | 0.7944 |
| 49.1  | 50    | 0.6778 | 35.2  | 35    | 0.7274 | 23.4  | 21    | 0.7853 | 16.0  | 16    | 0.8277 |
| 59.0  | 60    | 0.7333 | 41.0  | 40    | 0.7726 | 26.0  | 24    | 0.8147 | 17.5  | 18    | 0.8500 |
| 69.2  | 70    | 0.7796 | 46.3  | 45    | 0.8065 | 26.9  | 27    | 0.8235 | 18.7  | 20    | 0.8666 |
| 83.4  | 80    | 0.8296 | 55.1  | 55    | 0.8516 | 28.8  | 30    | 0.8412 | 21.8  | 22    | 0.9000 |
| 95.7  | 90    | 0.8630 | 60.3  | 65    | 0.8725 | 28.8  | 33    | 0.8412 | 25.4  | 24    | 0.9277 |
| 111.1 | 100   | 0.8954 | 68.6  | 75    | 0.9000 | 32.5  | 36    | 0.8706 | 26.3  | 26    | 0.9333 |

$$E \cdot t = \left( \frac{1}{k_2} - \frac{K_3}{k_1} - \frac{K_3 k_{-1}}{k_1 k_2} \right) \cdot a \cdot a - \left( \left( \frac{1}{k_1} + \frac{k_{-1}}{k_1 k_2} \right) (1 + K_3 \cdot a) \right) \log (1 - a)$$

where  $t$  is time,  $a$  is degree of reaction and  $A$  and  $B$  are constants. From graphs of  $t/a$  versus  $\log (1-a)/a$   $A$  and  $B$  are determined as the intercept on the ordinate and the slope, respectively. The constants  $A$  and  $B$  for four experiments with different initial concentrations of casein but with the same enzyme concentration are given in Table 1. In Table 2 the degrees of reaction are given together with measured and calculated values of  $t$  for the same four experiments.

As shown in Table 1 the constant  $A$  is negative,  $B$  is positive and both constants are proportional to the initial concentration of substrate. The tryptic action on casein thereby resembles that on  $\beta$ -lactoglobulin, and the mechanism suggested for this reaction<sup>1</sup> could hold for the action on casein too. A mechanism involving an inhibition by the split products also could lead to a chronometric integral identical with that found empirically and so be a possible mechanism for the action of trypsin on casein and  $\beta$ -lactoglobulin.

The mechanism:



gives the following chronometric integral by steady state treatment: (see eqn. above) which can give a negative coefficient to the zero order term and constants proportional to casein concentration if  $K_3(k_2 + k_{-1}) > k_1$  and  $K_3 \cdot a \gg 1$ . ( $X_n$  might be an enzyme form, a combination of enzyme and substrate or a combination of enzyme and products,  $C$  is casein,  $P$  is reaction products,  $E$  is enzyme concentration,  $k_n$  is velocity constant for the  $n$ 'th reaction and  $K_3$  is the equilibrium constant for the reaction ( $\pm 3$ )).

*Acknowledgement.* The authors thanks are due to professor dr D. Müller who kindly has permitted us to use the facilities of his laboratory. Financial support from *Statens Videnskabsfond, Købmand i Odense Johann og Hanne Weimann, født Seedorffs Legat* and *Carlsbergfondet* from Denmark, as well as the receipt of *Nordisk Forskningsstipendium* from the University of Oslo, Norway, is gratefully acknowledged by J. G.

1. Graae, J. *Biochim. et Biophys. Acta* **32** (1959) 510.
2. Graae, J. and Hansen, B. O. *Acta Chem. Scand.* **15** (1961). *In press.*
3. Kunitz, M. *J. Gen. Physiol.* **30** (1947) 291.

Received March 14, 1961.

## Über die Rotationsdispersion von Tris-Diamin-Chrom(III)-Komplexen

O. KLING\* und F. WOLDBYE

*Chemisches Laboratorium A, Die Dänische Technische Hochschule, Kopenhagen, Dänemark*

Bei der Messung der Absorptionsspektren von Chrom(III)-Komplexverbindungen wurde mehrfach festgestellt<sup>1-3</sup>, dass das bei etwa  $15\,000\text{ cm}^{-1}$  ( $\sim 650\text{ m}\mu$ ) liegende "Chromdublett", das auf Quartett-Dublett-Übergänge zurückzuführen ist<sup>4,5</sup>, eine ausgeprägte Feinstruktur zeigt. Der Zusammenhang dieser schwachen Interkombinationsbanden mit gewissen Normalschwingungen des Moleküls<sup>6</sup> legte es nahe zu untersuchen, ob in diesem Spektralgebiet auch Beziehungen zwischen spektraler Absorption und Rotationsdispersion optisch aktiver Chrom(III)-Komplexe bestehen.

Für eine erste Untersuchung dieser Frage wurden die Luteochelate  $[\text{Cr en}_3]^{3+}$  und  $[\text{Cr tn}_3]^{3+}$  ( $\text{en} = \text{Aethylendiamin}$ ,  $\text{tn} = \text{Trimethyldiamin}$ ) ausgewählt, da bei ihnen die schwachen Dublettbanden noch nicht von den ersten hohen Zentralionenbanden überdeckt werden sondern frei liegen<sup>1</sup>. Rotationsdispersionskurven dieser beiden Verbindungen im Wellenzahlgebiet  $\sim 17\,000 - 28\,000\text{ cm}^{-1}$  ( $\sim 600 - 350\text{ m}\mu$ ) sind bereits seit längerem bekannt<sup>6-8</sup>. Mit Hilfe der in diesem Institut entwickelten Messeinrichtung<sup>8,9</sup> war es nun möglich,

\* Derzeitige Anschrift: Institut für physikalische Chemie der Universität, Frankfurt a.M., Deutschland.