The Copper(II) Complexity of O-Phosphorylethanolamine

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The complex formation between copper(II) and O-phosphorylethanolamine (H2A) ions has been studied in 0.15 M KCl medium at

25° by pH titration.

By an adaption of Hedström's method 39 the concentration of free O-phosphorylethanolamine was calculated. The experimental data could be explained by formation of the complexes: CuHA+, CuA, Cu(HA)A and CuA². The corresponding equilibrium constants were computed (Table 2).

Probable coordination sites involved in the complexes are suggested, and are discussed in relation to copper(II) binding of phospho-

proteins and phosphatides.

 ${f A}$ s pointed out by several authors 1-5 phosphoproteins and phosphopeptides ${f A}$ may under physiological conditions act as strong metal binders. The great metal affinity of these compounds has been partly ascribed to the presence of phosphorus 2,4,6-9, which in view of recent investigations 2,10-18 occurs mainly as monophosphate esters linked through the hydroxyl groups of serine and threonine residues *. Informations of the metal binding power of phosphoproteins and their break-down products may, thus, be obtained from studies on O-phosphorylated model compounds. Apparent stability constants of some metal complexes of O-phosphorylserine have previously been reported ^{20,21}. The present paper describes the copper(II) complexity of another model compound, O-phosphorylethanolamine; this ligand itself being an important metabolite existing in living tissues both in free ^{22–29} and combined state ^{28–35}.

The complex formation was measured by pH titration ³⁶⁻³⁸ at 25° in 0.15 M KCl. In this case several titrations for different ligand concentrations were carried out to allow calculation of the free ligand concentration more directly from the experimental data by graphical integration (cf. Ref. 39), assuming the possible presence of any set of mononuclear equilibria including hydrogen and hydroxo copper(II) complexes of the ligand. An equation required for this calculation was derived by applying the equations formulated by Hedström 39.

^{*} Apart from the phosphomonoester bond, the occurrence of other linkages of phosphorus has been suggested for some phosphoproteins 19.

The existing complexes and corresponding overall equilibrium constants were evaluated graphically. The pK values of the ligand, determined under the same experimental conditions 40, were used in the computations.

SYMBOLS

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Α
             O-phosphorylethanolamine with all dissociable hydrogen removed,
            H_2N \cdot CH_2 \cdot CH_2 \cdot O \cdot P
             molar concentration of free A
a
b_{\sigma}
            apparent stability constant of the complex CuA<sub>a</sub><sup>(2-2q)+</sup> defined by
             b_q\!=\![\mathrm{CuA}_q^{(2-2q)\,+}]\ [\mathrm{CuA}_{q-1}^{(4-2q)\,+}]^{-1}\ [\mathrm{Cu}^{2}\!+]^{-1}
            apparent stability constant of the complex CuHA+, defined by b_{\text{CuHA}} = [\text{CuHA+}] [\text{HA-}]^{-1} [\text{Cu}^{2+}]^{-1}
bcu(HA)A apparent stability constant of the complex Cu(HA)A, defined by
            b_{\text{Cu(HA)A}} = [\text{Cu(HA)A}^-] [\text{CuHA}^+]^{-1} [\text{A}^2^-]^{-1} apparent stability constant of the complex CuCl+, defined analogical to b_q total molar concentration of O-phosphorylethanolamine, defined by eqn. 5
b_{CuCl}
C_{\mathbf{A}}
C_{\mathbf{Cu}}
             total molar concentration of copper(II) ions, defined by eqn. 4
C_{\mathbf{H}}
            total molar concentration of hydrogen ions, defined by eqn. 6
 C_{C1}
             total molar concentration of chloride ions
h
            molar concentration of free hydrogen ions
            defined by eqn. 21
            number of protons bound in the proton complex HiA
K_{\mathsf{CuHA}}
            apparent acid ionization constant of the complex CuHA+, defined by
             K_{\text{CuHA}} = h \text{ [CuA] [CuHA+]}^{-1}
Kcu(HA)A apparent acid ionization constant of the complex Cu(HA)A, defined by
            K_{\text{Cu(HA)A}} = h[\text{CuA}_3^2] [\text{Cu(HA)A}]^{-1} defined by eqn. 22
\mathbf{M}
            g moles/litre
p,q,r
            number of HA, A^2, OH groups bound in the complex Cu(HA)_pA_q(OH)^{(2-p-2q-r)+}
            respectively defined by eqn. 9
R
            defined by eqn. 10
S
Z
            ligand number, defined by eqn. 8
\beta_2 apparent complexity constant, = b_1b_2 \beta_{\text{Cu(HA)A}} apparent complexity constant, = b_{\text{CuHA}}b_{\text{Cu(HA)A}} \varkappa_{1,1}\rho_{,0q,r} apparent overall equilibrium constant defined by eqn. 2
            apparent complexity constant defined by eqn. 3 defined on page 478
x_{j_1}
\varphi
            molar concentration of the species indicated
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MATERIALS

O-Phosphorylethanolamine, prepared according to Baer and Stancer ⁴¹, was a generous gift from Ing. G. Fölsch of this institute. The data of analysis for the present preparation are to be found in Ref. ⁴⁰ Aqueous stock solutions of this compound were standardized by potentiometric titrations. When not used, these solutions had to be kept frozen in order to obtain satisfactory reproducibility.

Aqueous stock solutions of copper(II) chloride were prepared from the British Drug

Houses Ltd. (A.R.) product and was standardized by an ion exchange method ⁴². Carbonate-free 0.2557 M KOH, containing 0.15 M KCl was prepared and standardized

The other reagents used in this work were of analytical grade and were not purified further. Tripple distilled water and weighed-in pipettes were used throughout.

METHOD

The quantities, h and $C_{\rm H}$, were varied at each pH titration, while $C_{\rm Cu}$ and $C_{\rm A}$ were held practically constant. This was carried out by successive delivering KOH to the titration solution, recording pH after each addition. As recording instrument a Radiometer PHM 3j valve potentiometer, equipped with a Radiometer glass electrode G 102 B, and a calomel electrode K 100 was used. The activity coefficients were assumed to be constant in the presence of 0.15 M KCl. All measurements were carried out at 25.00 \pm 0.03°.

The pH meter was standardized with 0.05 M potassium biphthalate and 0.05 M borax as recommended by Hitchock et al 43 . The pH meter readings were transformed to values of $-\log h$ by a calculated activity coefficient 40 being 0.79 \pm 0.02. It was thought to apply in the pH range, 4-7, involved in the computations below. For routine calibration of the entire measuring system, acetic acid titrations were used, cf. Ref. 40 , the deviations never exceeding 0.01 pH units.

Assuming O-phosphorylethanolamine only to interact with copper(II) ions, when both protons are removed from the phosphoryl group, i.e. as HA- and A^{2-} , the complex equilibria can be written

$$Cu^{2+} + pH^{+} + (p+q)A^{2-} + rH_{2}O \rightleftharpoons Cu(HA)_{p}A_{q}(OH)_{r}^{(2-p-2q-r)+} + rH^{+}$$
(1)
$$(p = 0,1,2,...; q = 0,1,2,...; r = 0,1,2,...; p+q+r = 1,2,3,...;)$$

Since H_2O is approximately constant, the system is defined by the overall equilibrium constants * (all charges omitted in the following for the sake of clarity):

$$\varkappa_{1,1p,0q,r} = \frac{[\operatorname{Cu}(\operatorname{HA})_p A_q(\operatorname{OH})_r]}{[\operatorname{Cu}] h^{p-r} a^{p+q}}$$
 (2)

The quantities p,q, and r may attain any integer values given above, restricted by $(p+q+r)_{\max}$, which is equal to the maximum coordination number of copper(II). Occurrence of polynuclear complexes was assumed to be negligible with the use of low values for $C_{\mathbf{A}}$ and $C_{\mathbf{Cu}}$, with $C_{\mathbf{A}}$ in excess of $C_{\mathbf{Cu}}$.

Analogously to eqn. 2, constants for the formation of proton complexes are defined

$$\kappa_{j1} = \frac{[H_j A]}{h^j a}$$
(j = 1,2,3)

The values of these constants, used in this work, have been re-calculated from the pK values reported earlier ⁴⁰.

Due to presence of bulk electrolyte (0.15 M KCl), the species CuCl⁺ has also to be considered. The total content of copper(II), ligand and dissociable hydrogen will thus be given by (cf. Ref.³⁸)

^{*} The notation of these constants is simplified, when only one or two complex forming groups are bound to the central metal ion. Thus will, e.g., the constants corresponding to CuOH+ and CuA be written $\varkappa_{1,1}$ and $\varkappa_{1,01}$, respectively.

$$C_{\text{Cu}} = [\text{Cu}](1 + \sum_{\mu_{1,1p,0q,r}} h^{p-r} a^{p+q} + b_{\text{CuCl}}[\text{Cl}])$$
 (4)

$$C_{\mathbf{A}} = a + \sum_{\mathbf{p}} (p+q) \kappa_{\mathbf{1},\mathbf{1}p,\mathbf{0}q,\mathbf{r}} [\operatorname{Cu}] h^{p-\mathbf{r}} a^{p+q} + \sum_{\mathbf{p}} \kappa_{j\mathbf{1}} h^{j} a$$
 (5)

$$C_{\rm H} = h - [{\rm OH}] + \sum_{(p-r)\kappa_{1,1p,0q,r}} [{\rm Cu}] h^{p-r} a^{p+q} + \sum_{j\kappa_{j1}} i \kappa_{j1} h^{j} a$$
 (6)

The summations have been carried out in respect of all stated values of p,q and r. The value of $C_{\rm H}$ for each point of titration may be calculated from

$$C_{\mathbf{H}} = 2C_{\mathbf{A}} - [KOH] \tag{7}$$

The figure $b_{\text{Cucl}}[\text{Cl}]$ in eqn. 4 will be constant as $[\text{Cl}] \approx C_{\text{Cl}} = 0.15$, and [OH]in eqn. 6 negligible in the $-\log h$ range involved.

From eqns. 4 and 5 we obtain the ligand number defined thus (cf. Ref. 38)

$$Z = \frac{C_{A} - a - \sum_{C_{Cu}} \kappa_{j1} h^{j} a}{C_{Cu}} = \frac{\sum_{(p+q)\kappa_{1,1p,0q,r} h^{p-r} a^{p+q}} (p+q) \kappa_{1,1p,0q,r} h^{p-r} a^{p+q} + b_{Cucl}[Cl]}{1 + \sum_{(p+q)\kappa_{1,1p,0q,r} h^{p-r} a^{p+q} + b_{Cucl}[Cl]}$$
(8)

It follows from eqn. 8 that the constants $\varkappa_{1,1p,0q,r}$ can be determined, if Z is known as a function of h and a. As Z is given by eqn. 8, if h and a are known, we have, thus, first to compute a. This computation as well as the evaluation of p,q,r and $\varkappa_{1,1p,0q,r}$ will be described after the following section.

MEASUREMENTS

Solutions for titration of metal complexes were prepared by pipetting stock solutions of O-phosphorylethanolamine, copper(II) chloride, potassium chloride and aq. dest. in order to obtain the initial concentrations C_A , C_{Cu} and 0.15 M KCl. The titrations were continued to pH 10.5. All titration equipment was the same as that described 17. The content (initially 10 ml) of the titration vessel was stirred and freed from carbon dioxide

by bubbling nitrogen, pretreated as previously ¹⁷. Five titrations with different C_A were carried out in this manner covering the concentration ranges $C_A = (0.8-3) \times 10^{-3}$, $C_H = (0.8-6) \times 10^{-3}$ and $C_{Cu} = 7.8 \times 10^{-4}$. Each titration could be reproduced within 0.01 in pH. The data used for the calculations below

constituted a mean of at least two titrations.

At $-\log \approx 6.3$ the measuring solutions grew slightly opalescent, developing into a greenish-blue precipitate at $-\log h \approx 7$. Semi-quantitative analysis of the washed and dried precipitate, isolated from a solution of pH 7.8, showed the presence of about 4 % N and 16 % P. This apparently indicates the precipitate to be a copper(II)-O-phosphorylethanolamine compound. Due to this sparing solubility of the present system above $-\log h \approx 6.3$, the computations were not extended further than up to $-\log h = 6.10$.

The results of the measurements are shown in Fig. 1, where the titrations are repre-

sented as $-\log h(C_{\rm H})_{\rm CA}$ curves.

DETERMINATION OF THE FREE O-PHOSPHORYLETHANOLAMINE CONCENTRATION

No true indications about the kind of complexes formed could be obtained from usual titration curves (as for example in the Cu(II)-O-phosphorylserine

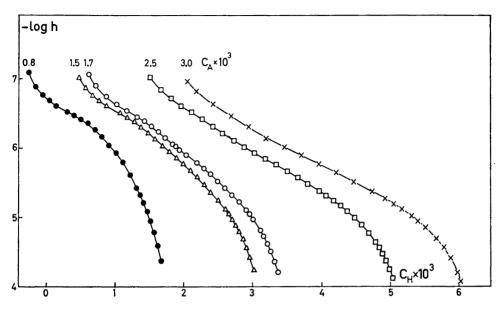


Fig. 1. The potentiometric measurements. $-\log h$ as a function of the total concentration of hydrogen ions, $C_{\rm H}$ (obtained from eqn. 7), for different total O-phosphorylethanolamine concentration, $C_{\rm A}$.

system ²¹). The free ligand concentration was therefore computed without any special assumptions about the complexes formed. For this purpose Hedström's method ³⁹ was made applicable to measurements and equilibria of the present type.

We define the functions R and S

$$R = \sum \varkappa_{j_1} h^j a = \sum [H_j A]$$
 (9)

$$S = \sum_{\kappa_{1,1}p,0q,r} h^{p-r} a^{p+q}$$
 (10)

Partial differentiation of R with respect to $\ln h$, followed by partial differentiation with respect to $\ln a$, results in

$$\left(\frac{\partial R}{\partial \ln h}\right)_a = \sum j \varkappa_{j_1} h^j a \tag{11}$$

$$\left(\frac{\partial^2 R}{\partial \ln h \ \partial \ln a}\right) = \sum j \varkappa_{j_1} h^j a = \left(\frac{\partial R}{\partial \ln h}\right)_a \tag{12}$$

Partial differentiation of S with respect to $\ln h$ and $\ln a$, followed by partial differentiation with respect to $\ln a$ and $\ln h$, gives

$$\left(\frac{\partial S}{\partial \ln h}\right)_{a} = \sum (p-r)\varkappa_{1,1p,0q,r}h^{p-r}a^{p+q}$$
(13)

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$$\left(\frac{\partial S}{\partial \ln a}\right)_{h} = \sum (p+q) \varkappa_{1,1} p_{,0q,r} h^{p-r} a^{p+q}$$
(14)

$$\left(\frac{\partial^2 S}{\partial \ln h} \frac{\partial}{\partial \ln a}\right) = \left(\frac{\partial^2 S}{\partial \ln a} \frac{\partial}{\partial \ln h}\right) = \sum_{i} (p-r)(p+q) \kappa_{1,1} p, q, r, h^{p-r} a^{p+q}$$
(15)

We may eliminate [Cu] from eqns. 5 and 6 by extension of the expression in those equations, containing [Cu] with C_{Cu} . By substituting eqns. 9-11 and 13-14 into the resulting equations, followed by partial differentiation with respect to $\ln h$ and $\ln a$, respectively, we obtain

$$\left(\frac{\partial C_{\mathbf{A}}}{\partial \ln h}\right)_{a} = C_{\mathbf{Cu}} \left[\frac{(1+S+b_{\mathbf{Cucl}}[\mathbf{Cl}])\left(\frac{\partial^{2}S}{\partial \ln a \ \partial \ln h}\right) - \left(\frac{\partial S}{\partial \ln a}\right)_{h}\left(\frac{\partial S}{\partial \ln h}\right)_{a}}{(1+S+b_{\mathbf{Cucl}}[\mathbf{Cl}])^{2}} \right] + \left(\frac{\partial R}{\partial \ln h}\right)_{a}$$
(16)

$$\left(\frac{\partial C_{\rm H}}{\partial \ln a}\right)_{h} = C_{\rm Cu} \left[\frac{(1+S+b_{\rm Cucl}[{\rm Cl}])\left(\frac{\partial^{2}S}{\partial \ln h}\partial \ln h\right) - \left(\frac{\partial S}{\partial \ln h}\right)_{a}\left(\frac{\partial S}{\partial \ln a}\right)_{h}}{(1+S+b_{\rm Cucl}[{\rm Cl}])^{2}} \right] + \left(\frac{\partial^{2}R}{\partial \ln h}\partial \ln a\right) (17)$$

From eqns. 12 and 15 follows that the right hand sides of eqns. 16 and 17 are identical. Consequently

$$\left(\frac{\partial C_{\mathbf{A}}}{\partial \ln h}\right)_{\mathbf{a}} = \left(\frac{\partial C_{\mathbf{H}}}{\partial \ln a}\right)_{\mathbf{h}} \tag{18}$$

This equation may be transformed 39 to

$$\left(\frac{\partial \ln a}{\partial \ln h}\right)_{C_{\mathbf{A}}} = -\left(\frac{\partial C_{\mathbf{H}}}{\partial C_{\mathbf{A}}}\right)_{k} \tag{19}$$

Solving for $-\log a$ within the range $-\log h_0$ to $-\log h$ results in

$$-\log a = -\left[\int_{-\log h_0}^{-\log h} \left(\frac{\partial C_{\mathbf{H}}}{\partial C_{\mathbf{A}}}\right)_{h} d(-\log h)\right]_{C_{\mathbf{A}}} + (-\log a_0)$$
 (20)

From eqn. 20 values of $-\log a_0/a$ were determined by graphical integration ³⁹. The $-\log h(C_{\rm H})_{C_{\rm A}}$ curves of Fig. 1 were thus cut for constant $-\log h$, and $C_{\rm H}$ drawn against $C_{\rm A}$ (Fig. 2a). The quantity $\left(\frac{\partial C_{\rm H}}{\partial C_{\rm A}}\right)_h$ was determined, and plotted against $-\log h$ (Fig. 2b). The area under the resulting curve from $-\log h = -\log h_0$ to $-\log h$ gave then $-\log a_0/a$. After the calculation of a_0 from

$$a_0 = C_A/(1 + \kappa_{11}h + \kappa_{21}h^2) \tag{5a}$$

at a $-\log h$ value (= $-\log h_0$), where no complex formation could be detected, the corresponding a values were obtained. The a values, determined in this manner, are listed in Table 1. For each a value determined the corresponding Z value was calculated; the result has been compiled in the same table.

In spite the graphical integration method ³⁹ by Hedström usually requires many values for the known variables, the rather few C_{A} values of this study, seem sufficient, due to the constancy of $\left(\frac{\partial C_{H}}{\partial C_{A}}\right)_{h}$ in the C_{A} range involved (Fig. 2a).

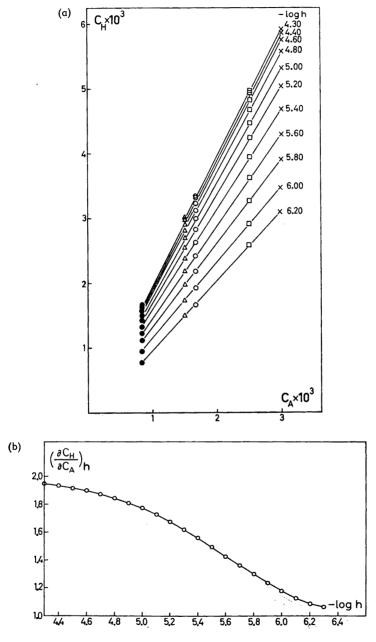


Fig. 2. Evaluation of the free O-phosphorylethanolamine concentration. (a). $C_{\rm H}$ vs. $C_{\rm A}$ for different $-{\rm log}h$. (b). $\left(\frac{\partial C_{\rm H}}{\partial C_{\rm A}}\right)_h$ vs. $-{\rm log}h$ for $C_{\rm A}=(0.8-3)\times 10^{-3}$.

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Table 1. Corresponding values of $-\log h$, $C_{\rm A}$, $C_{\rm H}$ (obtained from $-\log h(C_{\rm H})C_{\rm A}$ curves), a (obtained from eqns. 20 and 5a), and Z (from eqn. 8). $C_{\rm Cu} = 7.84 \times 10^{-4}$ $\varkappa_{11} = 1.259 \times 10^{10} \ {\rm M}^{-1}, \qquad \varkappa_{21} = 4.678 \times 10^{15} \ {\rm M}^{-2}$

	$ \begin{array}{c c} * & \times \\ C_{\mathbf{A}} = 3.010 \times 10^{-3} \\ -\log h_0 = 4.30 \end{array} $			$\begin{array}{c} & \square \\ C_{\rm A} = 2.519 \times 10^{-3} \\ -{\rm log}h_{\rm 0} = 4.30 \end{array}$			$\begin{array}{c} \bullet & \bullet \\ C_{\rm A} = 1.682 \times 10^{-3} \\ -\log h_{\rm 0} = 4.30 \end{array}$			$\begin{array}{c} * & \triangle \\ C_{\mathbf{A}} = 1.507 \times 10^{-3} \\ -\log h_{0} = 4.30 \end{array}$			$C_{A}^{*} = 0.834 \times 10^{-8}$ $-\log h_{0} = 4.40$		
-logh	$C_{ m H} imes 10^{ m 3}$	« 10°	Z	$C_{ m H} imes 10^3$	$ imes 10^9$	Z	$C_{ m H} imes 10^3$	<i>a</i> × 10 ⁹	Z	$C_{ m H} imes 10^3$	<i>a</i> × 10⁰	Z	$C_{\mathbf{H}} \times 10^3$	a × 109	Z
5.400 5.500 5.600 5.700 5.800 5.900	4.687 4.498 4.300 4.105 3.904 3.703	23.31 33.13 46.36 63.92 86.83 116.3	0.1261 0.1596 0.1985 0.2455 0.2974 0.3564	3.927 3.766 3.605 3.435 3.263 3.079	53.49 72.68 97.31	0.1076 0.1354 0.1688 0.2081 0.2528 0.3019	2.620 2.518 2.410 2.299 2.175 2.041	13.035 18.52 25.92 35.73 48.55	0.1427	2.366 2.270 2.177 2.071 1.965	11.67 16.59 23.215 32.01 43.48	0.0671 0.0841 0.1044 0.1285 0.1555	1.324 1.273 1.224 1.169 1.109	6.464 9.185 12.855 17.725 24.075	0.0381 0.0478 0.0591 0.0728
6.000	3.479	153.6	0.4227	2.903	J 28.55 167.65	0.3583	1.907		$0.2431 \\ 0.2837$		76.91	0.2194	0.948	42.59	

^{*} Initial concentration.

EVALUATION OF THE EQUILIBRIUM CONSTANTS

As can be seen from Table 1 all values of Z are below 1. To include all possibly existing complexes, $(p+q)_{\max}$ was therefore in the first place assumed to equal 2. For p=q=0, r was assigned to 1, but otherwise r was set equal to zero. Eqn. 8 will thus be reduced to

$$\frac{Z}{(1-Z)a} \varphi = \kappa_{1,11}h + \kappa_{1,01} + \frac{(2-Z)a}{(1-Z)} (\kappa_{1,12}h^2 + \kappa_{1,11,01}h + \kappa_{1,02})$$
(8a)

where $\varphi = 1 + \varkappa_{1,1}h^{-1} + b_{\text{CuCl}}[\text{Cl}]$

Since the term $b_{\text{Cucl}}[\text{Cl}]$ is approximately constant in this study (cf. p. 474), the expression φ in eqn. 8a will be constant for constant h. Varying values of $\varkappa_{1,1}$ in this expression are reported in the literature ⁴⁴. In the present study use will therefore be made of a range of values from $10^{-7.5}$ to $10^{-8.25}$ M. From these values follow that the amount of CuOH^+ will be rather negligible in the present $-\log h$ range.

To identify complexes formed, $Z(1-Z)^{-1}a^{-1}$ was first plotted against h (Fig. 3). In this plot of eqn. 8a the presence of CuHA⁺ and CuA only should give a straight line independant of C_A ; existing CuHA⁺ species making the ordinata values to decrease with h. At the occurrence of higher complexes, *i.e.* (p+q)>1, the plotted data should vary with C_A . As the latter case applies to Fig. 3, we may identify the higher complexes apparently present.

In Fig. 4 the quantity $Z(1-Z)^{-1}a^{-1}$ has therefore been plotted against $(2-Z)a(1-Z)^{-1}$ for constant h. As shown in the figure data are best represented by a serie of straight lines, indicating the true value of $(p+q)_{\max}$ to be 2. Conse-

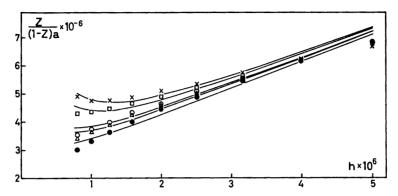


Fig. 3. $Z(1-Z)^{-1}a^{-1}$ vs. h for different C_A (eqn. 8 a). The curves are calculated for $\varkappa_{1,11}=1.1\times 10^{12}$ M⁻², $\varkappa_{1,01}=2.45\times 10^{6}$ M⁻¹, $\varkappa_{1,11,01}=2.3\times 10^{18}$ M⁻², $\varkappa_{1,02}=2.45\times 10^{12}$ M⁻²; $\varkappa_{1,1}=10^{-7,73}$ M, $b_{\text{CuCl}}=1.0$ M⁻¹. The symbols of the plotted points are those indicated in Table 1.

quently we may consider the probable existence of $Cu(HA)_2$, $Cu(HA)A^-$ and CuA_2^{-1} apart from $CuHA^+$ and CuA. In Fig. 4 there is, however, a slight upward trend for increasing C_A at high $-\log h$. For this reason the plotted points corresponding to $C_A > 2.52 \times 10^{-3}$ and $-\log h > 6.0$ were excluded at the calculation of slopes and intercepts of the straight lines.

From these intercepts (I) and slopes (L) the following functions were formed:

$$I\varphi = \varkappa_{1,11}h + \varkappa_{1,01} \tag{21}$$

$$L\varphi = \kappa_{1,12}h^2 + \kappa_{1,11,01}h + \kappa_{1,02} \tag{22}$$

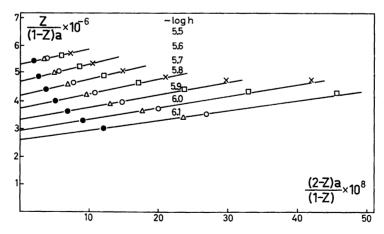


Fig. 4. $Z(1-Z)^{-1}a^{-1}$ as a function of $(2-Z)a(1-Z)^{-1}$ with h as parameter (eqn. 8a). The symbols are the same as indicated in Table 1.

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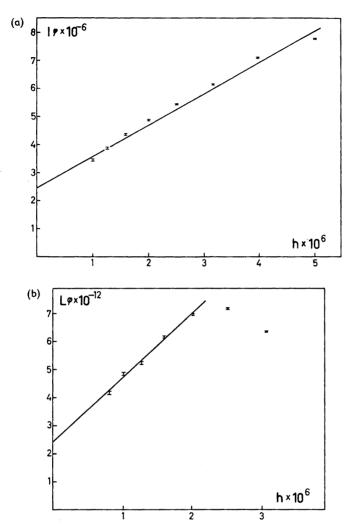


Fig. 5. Graphical evaluation of the overall equilibrium constants, assuming $\kappa_{1,1}$ (a) Eqn. 21 plotted as $L\varphi$ vs. h.

In Fig. 5a, eqn. 21 is plotted as the curve $I\varphi(h)$. For each h-value the abovementioned range of $\kappa_{1,1}$ is indicated (mean value $10^{-7.73}$ M). The constant b_{Cucl} was set equal to 1.0 M⁻¹ (Ref.⁴⁴). As shown in the figure, data can be fitted to a straight line with finite slope and intercept. The species CuHA+ and CuA are thus obviously existing. The intercept $(\varkappa_{1,01})$ and the slope $(\varkappa_{1,11})$ of the best straight line furnished the equilibrium constants:

$$egin{aligned} & \pmb{\varkappa_{1,01}} = (2.45 \pm 0.2) \times 10^6 \, \mathrm{M^{-1}} \\ & \pmb{\varkappa_{1,11}} = (1.1 \, \pm 0.2) \times 10^{12} \, \mathrm{M^{-2}} \end{aligned}$$

In a similar manner $L\varphi$ has been plotted against h in Fig. 5b. Even in this case, data can be fitted to a straight line, which may provide evidence of existence of the species $Cu(HA)A^-$ and CuA_2^{2-} . Formation of $Cu(HA)_2$, in measurable quantities can reasonable be ruled out. In the present plot data should then have been represented as a parabola with its vertex downwards. The values of equilibrium constants related to $Cu(HA)A^-$ and CuA_2^{2-} were computed from the best straight line to be:

$$\varkappa_{1,02} = (2.45 \pm 0.2) \times 10^{12} \,\mathrm{M}^{-2} \\
 \varkappa_{1,11,01} = (2.3 \,\pm 0.2) \times 10^{18} \,\mathrm{M}^{-3}$$

The logarithms of the computed overall equilibrium constants, $\varkappa_{1,1p,0q,r}$, were converted into logarithms of apparent stability constants and apparent complexity constants of the complexes, as well as into pK values of the hydrogen copper(II) complexes. This is evident from Table 2, where all these constants are given in logarithmic form.

DISCUSSION

As can be seen in Fig. 3, the curves calculated from the computed constants reasonably agree with the plotted data. As expected the deviations are most apparent at high h, as the pH decrements due to complex formation are least pronounced there.

The complexes evaluated as formed in this study, *i. e.* CuHA⁺, CuA, Cu(HA)A⁻ and CuA²₂, may not exclude the existence of other complexes for true, and at least some other possibilities may therefore be discussed.

An indication of the existence of complexes with more than two O-phosphorylethanolamine groups per central ion, thus, seems to be the upward trend of the curves of Fig. 4 at high —logh. This trend is, however, too slight to allow any accurate proof of the existence of such species.

In the present computations, only a hydroxo complex $CuOH^+$ was considered, which apparently is the only inorganic hydroxo species formed in the present concentration ranges ⁴⁴. Other monohydroxo complexes, *i. e.* in the first place Cu(HA)OH and $CuA(OH)^-$, would at $-\log h = 6.1$ (the upper limit of $-\log h$ used in this work) apparently constitute less than 5 % of the corresponding species $CuHA^+$ and CuA, cf. Ref. ⁴⁵. These hydroxo complexes, consequently, cannot appreciably change the proposed reaction scheme. This is also evident from the curves of Figs. 3 and 5a, as the occurrence of a large amount of $CuA(OH)^-$ would have been indicated as an upward trend for decreasing h. The right hand side of eqn. 8a should then include the figure $\varkappa_{1,01,1}h^{-1}$. The presence of Cu(HA)OH on the other hand, cannot be excluded, as these measurements do not distinguish this complex from CuA.

The apparent negligible amount of these mononuclear hydroxo species may exclude the existence of ol complexes $^{7,46-50}$. Polynuclear complexes, where one copper(II) ion is coordinated through either the phosphoryl and amino group of the ligand, can reasonably also be excluded. In the $-\log h$ range involved the pK value of the ligand amino group (=10.10) does not allow for such complex formation due to the relatively low copper(II)-ammine affinity (see, e. g., $\log b_1$ for the Cu(II)-ammine system 44), cf. Ref. 51

Table 2. Logarithms of $\varkappa_{1,1p,0q,r}$ constants, apparent stability constants, and apparent complexity constants of the complexes, and apparent pK values of the acid complexes in 0.15 M KCl medium at 25°.

$$\begin{array}{lll} C_{\rm A} = (0.8-3)\times 10^{-3}, & C_{\rm Cu} = 7.84\times 10^{-4}, & -{\rm log}h = 5.30-6.10. \\ {\rm log}\varkappa_{11} = 10.10, & {\rm log}\varkappa_{21} = 15.67, & \varkappa_{1,1} = 10^{-7.73}{\rm M}, & b_{\rm CuCl} = 1.0~{\rm M}^{-1} \end{array}$$

Re	actic	on	Log. eq			
$Cu^{2+} + H^{+} + A^{2-}$	₽	CuHA+		$\log \varkappa_{1,11}$	=	12.04
Cu ²⁺ + HA	~`	CuHA+	$\log \frac{\varkappa_{1,11}}{\varkappa_{11}}$	$=\log b_{\mathrm{CuHA}}$	=	1.94
Cu ²⁺ + A ²⁻	€3	CuA	$\log_{\varkappa_{1,01}}$	$=\log b_1$	=	6.39
$Cu^{2+} + H^{+} + 2A^{2-}$	/	Cu(HA)A		$\log \varkappa_{1,11,01}$	=	18.36
$Cu^{2+} + HA^{-} + A^{2-}$	=	Cu(HA)A~	$\log \frac{\varkappa_{1,11,01}}{\varkappa_{11}}$	$=\log\!eta_{\mathrm{Cu}(\mathrm{HA})\mathrm{A}}$	=	8.26
CuHA + A2-	72	Cu(HA)A	$\log \frac{\varkappa_{1,11,01}}{\varkappa_{1,11}}$	$=\log b_{\mathrm{Cu(HA)A}}$	==	6.32
Cu ²⁺ + 2A ²⁻	=	CuA2-	$\log_{\mathcal{U}_{1,02}}$	$=\log\!eta_2$	=	12.39
CuA + A2-	=	CuA2~	$\log \frac{\varkappa_{1,02}}{\varkappa_{1,01}}$	$=\log b_{\mathbf{z}}$		6.00
CuHA+	47	$CuA + H^+$	$\log \frac{\varkappa_{1,11}}{\varkappa_{1,01}}$	$= pK_{CuHA}$	=	5.65
Cu(HA)A-	⇌	$\mathrm{CuA_{2}^{2-}+H^{+}}$	$\log \frac{\varkappa_{1,11,\theta_1}}{\varkappa_{1,02}}$	$= pK_{Cu(HA)A}$	100	5.97

The existence of $CuHA^+$ and $Cu(HA)A^-$ provides evidence for the ligand phosphoryl group to act complexing. This is in contrast to the Ag(I) — Ophosphorylethanolamine system 52 , where the species AgA^- and AgA_2^{3-} only could be detected in a similar concentration range as the present. The amino group of the ligand was suggested as single coordination site for the latter species 52 , which may be attributed to Ag(I) preferably forming linear complexes with this type of ligand, cf. Ref. 53 . The great difference between b_{CuHA} and b_1 in this work (cf. Table 2) indicates the ligand amino group to be introduced as dentate site, when $CuHA^+$ is converted to CuA. The species CuA and CuA_2^{2-} may, thus, be coordinated through the ligand phosphoryl and amino group or the amino group only. The former alternative is more probable, at least for CuA, as $logb_1$ of the Cu(II)-ammine system 44 is reported to be about 2 log. units lower than that of the present. The effect of the increased net negative charge, caused by the presence of the phosphoryl group, cannot reasonable alone explain this great difference in b_1 . — Due to the same fact

copper(II) reasonably interacts with the phosphoryl group also in the corresponding O-phosphorylserine 1:1 complex 21. The central copper(II) ion is, however, in this complex apparently mainly coordinated through a fivemembered chelate ring formed by the amino and carboxyl donor groups. The value of $\log b_1$ (= 9.64) in this system is about 1.5 - 2 log. units greater than logb₁, in copper(II) systems of similar unphosphorylated amino acids ^{38,54,55}.

Klotz et $al.^{6,56}$ studied the copper(II) binding properties of α - and β -casein. The a-case was found to have the greater copper (II) binding capacity, being greater than that of any other protein investigated 6. The difference in copper (II) affinity of α - and β -casein paralleled the phosphorus content of those proteins, suggested to be due to difference in net negative charge 6. The apparent complexing action of the phosphoryl group in the present complexes, and in the O-phosphorylserine complexes ²¹ may indicate that the greater copper(II) binding power of these phosphoproteins is partly due to complex formation through phosphoryl groups.

In the phospholipids, phosphatidylethanolamine and phosphatidylserine, where the phosphoryl group is diesterified, the same groups are free to react as those proposed as main dentate sites in O-phosphorylethanolamine and O-phosphorylserine. Since the pK values of the amino group in these phosphatides are of the same size 57 as those of the corresponding low molecular weight phosphates 21,40, both types of compounds may at a physiological pH form copper(II) complexes of nearly the same strength. The complexes formed by those ligands will then be those, where the dissociable proton is removed from the amino nitrogen group due to the pK value of the hydrogen copper(II) complexes (cf. Table 2 and Ref.²¹). This gives evidence for a considerable copper(II) complexity of these phosphatides, which may have biological significance. At inhibition of tromboplastinase action by copper(II) ions 58, for instance, complexes will obviously be formed between Cu(II) ions and the tromboplastical active lipid substrate, mainly constituting phosphatidylethanolamine and phosphatidylserine 59.

Acknowledgements. The author wishes to express his gratitude to Docent Ido Leden for valuable discussions. The skilful technical assistance of Mr. Åke Lundquist is greatly acknowledged. The investigation was aided by grants from the Medical Faculty, University of Göteborg.

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Received October 7, 1959.