Haemoglobin Present in the Nuclear Fraction of the Liver

R. BONNICHSEN, G. EHRENSTEIN, G. HEVESY and J. SCHLIACK

Institute for Organic Chemistry and Biochemistry, University of Stockholm, Sweden

The nuclear fraction of 1 g of rat and rabbit liver contains 0.34 and 0.58 mg of in aqueous medium non-extractable haemoglobin. The corresponding figure for the nuclei of the red corpuscles of 1 ml of hen's blood is 0.9 mg.

The haemoglobin content of the nuclear fraction of 1 g of foetal rabbit liver is 100 times as large as that of 1 g of maternal liver. The ratio of their ⁵⁹Fe content 11 to 17 h after labelling the maternal plasma iron, works out to be 100.

One day or more after exposure of the rat to 500 r, incorporation of ⁵⁹Fe into the non-extractable haemoglobin of the liver nucleus fraction is strongly depressed.

As mentioned in a previous note (Bonnichsen et al.¹) the nuclear fraction of the liver of the guinea-pig contains small amounts of haemoglobin not extractable by saline or other aqueous solutions. This paper contains data on the incorporation of ⁵⁹Fe into this fraction isolated from the liver of adult rats and rabbits and of that of the rabbit embryo. This fraction was furthermore located in the nuclei of the erythrocytes of the hen.

That in the basophilic erythroblasts of human and rat bone marrow the site of haemoglobin synthesis is primarily the nucleus, was repeatedly suggested. The methods applied in these investigations were ultraviolet absorption microspectroscopy and cytochemical staining procedures ^{2–9}.

EXPERIMENTAL

In 15 experiments with male rats an aggregate number of 173 animals, weighing between 150–280 g were injected intraperitoneally with 0.25 ml of a 3.8 % ammonium-citrate solution containing 0.5–12 μ g of with 59 Fe labelled iron of 0.1–12 μ C activity. Half the number of rats was exposed to 150 r – 500 r of X-rays. Injection took place 15 min to 5 days after exposure.

The animals were killed from 2 to 48 h after injection. The livers were perfused first with 0.145 M NaCl and then with 0.25 M sucrose containing 0.018 m CaCl₂. The weighed livers were homogenized in 9 vols. of 0.25 M sucrose - 0.0018 M CaCl₂. The further procedure was carried out according to Hogeboom *et al.*¹⁰

The nuclear fraction isolated by this procedure was cytologically inhomogeneous. The fraction contained about 60-80 % of the cell nuclei, it also contained erythrocytes, connective tissue, residual intact cells and free mitochondria.

In order to haemolyze the erythrocytes, the fraction was homogenized with 10 vols. distilled water and allowed to stand several hours. This last step was repeated at least three times more.

After clearing the suspension of nuclei obtained in this way by adding 3 % of deoxycholate, the haemoglobin bands were steadily visible in the handspectroscope. The COband was that of haemoglobin. The pyridine-haemochromogen band was located at

The haemin was extracted with a mixture of acetone and HCl (10 ml 20 % HCl in 11 of acetone). After filtration, the acetone was evaporated in vacuo. The haemin was crystallized twice from conc. acetic acid, and the crystals washed with 1 N HCl. The haemin was then combusted and the solution analyzed as described by Bonnichsen et al.11

In order to know in which part of the above nuclear fraction the non-extractable haemoglobin is located, we separated the fraction, prior to haemolyzing the red cells with distilled water, in the counter-streaming centrifuge of Lindahl 12 in five fractions containing particles of different size and different specific gravity. The fractions were examined with the phase-contrast microscope and with the handspectroscope. Thereafter, the specific activity of their haemin was determined. The result of one of these experiments is seen in Fig. 1.

In five of the above experiments of the liver nuclei have been isolated both in aqueous medium and in organic solvents of low polarity according to the methods described by Dounce et al.¹³ and Allfrey et al.¹⁴ The nuclear preparations obtained in this way were treated and analysed as described above. No difference was found in the properties of the non-extractable haemoglobin of the nuclear fraction prepared either in aqueous or in organic medium.

The liver ferritin was prepared as previously described by Loftfield et al.¹⁵
In experiments with 2-3 kg rabbits 8 animals were investigated. Their plasma was labelled with ⁵⁹Fe as described by Ehrenstein et al.¹⁶ and reinjected. The animals were killed 3-24 h after injection.

Simultaneously, in all our experiments haemin of the circulating haemoglobin was analysed as well.

In two experiments with hens 100 ml of hen blood were incubated in vitro for 3 h at 37° C with 2 ml of 3.8 % ammonium citrate solution containing $2~\mu\mathrm{g}$ labelled iron of $25~\mu\mathrm{C}$ activity. The plasma was centrifuged off and the red cells washed four times with 5 vols. or more of isotonic saline.

The erythrocyte nuclei were prepared according to Hogeboom et al. 10 However, as the red cells are not sufficiently broken down by the homogenization procedure usually employed for the disintegration of other tissues, we haemolyzed the cells by freezing at -20° C and thawing them three times or more, suspended in a mixture of 0.25 M sucrose — 0.00018 m CaCl₂. The nuclei were then centrifuged down (International Refrigerated centrifuge head No. 269) for 10 min at 2 000 r.p.m. Thereafter, the procedure of Hogeboom et al.10 was applied. Samples were taken from the whole washed red cells, the stroma-free haemolysate and from the final nucleus preparation. The haemin was extracted as described above.

RESULTS AND DISCUSSION

a) Experiments with rats.

The amount of 59 Fe present in one μg of iron of the nuclear haemoglobin fraction of the liver varied in experiments on rats between 0.6×10^{-4} and 10×10^{-4} % of that injected.

The liver of our rats contained 10 μ g of nuclear haemin iron out of 2.5 mg of total iron present in the liver. This fraction makes out 0.4 % of its total iron content.

In the course of the purification process in aqueous medium extractable haemoglobin may have been removed. However, the non-extractable haemoglobin content of the nuclei was not found to be larger when the nuclei were

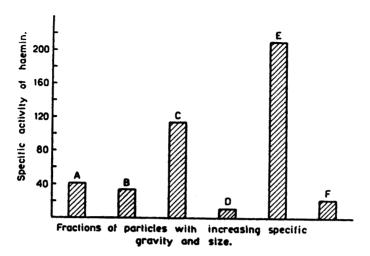


Fig. 1. Results of separation of nuclear components of the liver cells of the rat in Lindahls counter-streaming centrifuge.

A = Total nuclear fraction, prepared according to Hogeboom.

B = Impure mitochondria containing erythrocytes.

C = Erythrocytes and small fibers of connective tissue.

D = Cell membranes.

E = Small nuclei.

F = Large nuclei.

isolated after lyophilisation in organic medium. In the haemoglobin content of the nuclear fraction of the red cells of the hen, when these were isolated in an organic medium, Stern $et\ al.^{17}$ found a haemoglobin-iron content of 19 % of that of the total red corpuscle, while we find after isolation of the nuclei in an aqueous medium 1 % of the total haemoglobin-iron content to be non-extractable. This 1 % compares with 0.4 % found by us in the liver nuclei of rats and 2 % in those of the rabbit.

We compared also the effect of irradiation with 500 r of X-rays on rats on the incorporation of 59 Fe into the liver nuclear haemoglobin fraction. As seen in Table 1 the X-ray effect is shortly after exposure a very restricted one for both haemoglobin fractions. The X-ray effect as seen in Fig. 2 doesn't seem as pronounced as that on the circulating haemoglobin. If we, however, take into account the enhanced activity of the liver of the exposed animal, this difference is strongly reduced.

b) Experiments with rabbits.

The amount of ⁵⁹Fe present 4 h after injection in one μ g iron of the non-extractable haemoglobin of the nuclear fraction of the rabbit liver varied between 2 \times 10⁻⁴ and 6 \times 10⁻⁴ % of that injected.

The iron content of the non-extractable liver nucleus fraction haemo-globin varied between 0.5 and 3.8 μg with a mean value of 1.9 μg per one g liver.

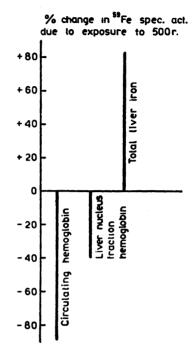


Fig. 2. Effect of exposure of the rat to whole body irradiation on the incorporation of intraperitoneally injected ⁵⁹Fe as citrate into liver fractions and the circulating haemoglobin.

For the nuclear haemoglobin content of 1 g of rat and rabbit liver the corresponding figures are 0.38 mg and 0.56 mg.

We also carried out experiments with pregnant rabbits 21 days after mating. A plasma sample of the mother was labelled as described by Ehrenstein *et al.*¹⁶ The livers of the mothers and foetuses were investigated 11 to 17 h after labelling the maternal plasma. The results are seen in Table 2.

Table 1. Ratio of specific activities of iron fractions of to 500 r exposed and control rats.

Injected with ⁵⁹ Fe shortly after exposure			Injected with ⁵⁹ Fe 1 day or more after exposure		
Liver nucleus fraction haemoglobin	Circulating haemoglobin	Total liver iron	Liver nucleus fraction haemoglobin	Circulating haemoglobin	Total liver iron
1.16 0.97 1.15 0.78	0.53 1.04 0.84 0.46	1.60 1.41 0.93 1.27	0.915 0.503 0.570 0.727 0.318	0.161 0.063 0.238 0.072 0.029	1.15 2.48 1.72 1.92 2.44
Mean value 1.01	0.72	1.30	0.605	0.112	1.94

Acta Chem. Scand. 11 (1957) No. 1

Table 2.

	Mother	Foetus
Total liver iron Total non-extractable liver nuclear haemoglobin iron Total liver activity in % of that injected Total non-extractable liver nuclear haemoglobin	13.9 mg 24 µg 8	$\begin{array}{c} 1.5~\mathrm{mg} \\ 41~\mu\mathrm{g} \\ 20 \end{array}$
activity in % of that injected	3.4×10^{-3}	3.4×10^{-1}

Both the iron content of the nuclear fraction and its specific activity is much higher in the foetus than in the mother.

Three of the pregnant rabbits were exposed 1 day prior to injecting them with 59 Fe to an X-ray dose of 500 r. The effect of the radiation on the incorporation of ⁵⁹Fe into the liver nuclear fraction is discussed in a paper by Ehrenstein et al.16

The generous support to this investigation by Riksföreningen för kräftsjukdomarnas bekämpande, Cancernämnden and the Alice och Knut Wallenbergs Stiftelse is gratefully acknowledged. We are also much indebted to Professor P. E. Lindahl for kindly putting his centrifuge at our disposal.

REFERENCES

- 1. Bonnichsen, R. and Hevesy, G. Acta Chem. Scand. 9 (1955) 1045.

- Villa, L. Haematologica 10 (1929) 327.
 Villa, L. Haematologica 10 (1929) 355.
 Riecker, H. H. J. Exptl. Med. 49 (1929) 937.
- 5. Knoll, W. Folia Haematol. 44 (1931) 310.
- Hermansky, F. Casopis Lékaru. Ceskych. 90 (1951) 988.
 De Carvalho, S. and Wilkins, M. F. H. Proc. 4th Internat. Congr. Int. Soc. Haematol., Grune & Stratton, New York, 1954, p. 119. 8. De Carvalho, S. Ed. Gaz. Med. Portug., Lisbon, 1954.
- 9. De Carvalho, S. Blood 10 (1955) 453.
- 10. Hogeboom, G. H., Schneider, W. C. and Striebich, M. J. J. Biol. Chem. 196 (1952) 111.

- Bonnichsen, R. and Hevesy, G. Acta Chem. Scand. 9 (1955) 509.
 Lindahl, P. E. and Nyberg, E. Iva 26 (1955) 309.
 Dounce, A. L., Tishkoff, G. H., Barnett, S. R. and Freer, R. M. J. Gen. Physiol. 33 (1950) 629.
- 14. Allfrey, V. G., Stern, H., Mirsky, A. E. and Saetren, H. J. Gen. Physiol. 35 (1952)
- Loftield, R. B. and Bonnichsen, R. Acta Chem. Scand. 10 (1956) 1547.
 Ehrenstein, G. and Hevesy, G. Acta Physiol. Scand. 38 (1956) 184.
- 17. Stern, H., Allfrey, V. G., Mirsky, A. E. and Saetren, H. J. Gen. Physiol. 35 (1952) 559.

Received October 15, 1956.