

(1 g) was then added with vigorous shaking. A precipitate immediately separated. After standing for 24 hours, the yellow precipitate was filtered off, washed and dried. Yield: 2.6 g (calc. 2.8 g). The compound was dissolved in 100 ml boiling chloroform, the solution filtered and cooled in ice. Orange-red (dichromate-coloured) crystals separated. The compound was recrystallised once more from chloroform. It does not melt on heating, but gradually turns black without change of the crystal form. Found: C 17.59; H 1.59; Pt 34.97. Calc. for  $[\text{PtI}_2(\text{C}_8\text{H}_8)]_n$ , (553.2): C 17.37; H 1.46; Pt 35.38.

Molecular weight (cryoscopically in bromoform solution): 492, 610. Dipole moment:

$c = 0.00198$  molar,  $\Delta\epsilon = 0.0130$ ,  $\mu = 7.3\text{D}$ ;  
 $c = 0.00406$  molar,  $\Delta\epsilon = 0.0240$ ,  $\mu = 7.0\text{D}$ .

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### Release of Hydantoins from Proteins \*

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In a note by the author<sup>1</sup>, the procedure of Fraenkel-Conrat and Fraenkel-Conrat<sup>2</sup> for the release of hydantoins from proteins was erroneously quoted, the temperature for the reaction being given at 75° C instead of 36° C (cf. Fraenkel-Conrat and Fraenkel-Conrat<sup>3</sup>).

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\* Correction to Attempted Successive Applications of the Edman Degradation to Insulin<sup>1</sup>.

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### The Occurrence in Lichens of the Folic Acid-, Folinic Acid-, and Vitamin B<sub>12</sub>-Group of Factors

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In connection with investigations on the folic acid-, folinic acid-, and vitamin B<sub>12</sub>-group of factors in algae it was considered of interest to study the occurrence of these factors in some lichens, a group of plants known to represent a symbiosis between fungi and algae. The following lichens have been studied using microbiological and bioautographic methods: *Cladonia silvatica*, *Umbilicaria pustulata*, *Parmelia physodes*, *Parmelia furfuracea*, *Cetraria islandica*, *Evernia prunastri*, *Alectoria jubata*, and *Usnea comosa*. The lichens were collected at Grönvik, Nämndö.

*Escherichia coli* 113-3<sup>1</sup> served as a test organism for the vitamin B<sub>12</sub>-factors, *Leuconostoc citrovorum* ATCC 8081<sup>2</sup> for the folinic acid and *Streptococcus faecalis* ATCC 8043 for the folic acid tests. The organisms were utilized in the agar cup plate method. For *Streptococcus faecalis* the Difco folic acid assay medium with 1.6 % Bacto agar was used<sup>3</sup>. The solvent for the chromatographic separation was sec. butanol saturated with water, and containing 3 % acetic acid and 25 mg KCN/l.

The lichens were carefully cleaned and dried at room temperature. Two grammes of finely ground material were suspended in 25 ml of water or buffer solution. Three different methods of freeing the active substances were tried, namely, extraction with water at 37° C for 24 hrs, autoclaving for 20 min. at 120° C in water containing small amounts of KCN, and enzymatic treatment with a chicken pancreas homogenate at 37° C for 24 hrs at pH 7.5. Autoclaving gave the highest

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Table 1. Total activity expressed in  $\mu\text{g}$  standard per g dry weight of lichens.

Lichenes *	<i>E. coli</i> Vit. B <sub>12</sub> std.	<i>L. citrovorum</i> Leucovorin std	<i>S. faecalis</i> Folvite std.
<i>Cladonia silvatica</i>	0.035	0.68	0.26
<i>Umbilicaria pustulata</i>	0.02	0.19	0.24
<i>Parmelia furfuracea</i>	< 0.005	0.09	0.43
<i>Parmelia physodes</i>	0.02	2.35	1.16
<i>Cetraria islandica</i>	< 0.005	1.80	1.16
<i>Evernia prunastri</i>	0.008	0.06	0.26
<i>Alectoria jubata</i>	< 0.005	0.01	0.41
<i>Usnea comosa</i>	0.015	0.28	0.41

\* Symbiosis of fungi belonging to *Discomycetes*, order *Lecanorales* and algae belonging to *Chlorophyceae*, order *Chlorococcales*.

values for *E. coli*, enzymatic treatment the highest values for *L. citrovorum* and *S. faecalis*.

The results are given in Table 1.

Table 2 shows the various types of growth factors for *S. faecalis* and *L. citrovorum* as revealed after chromatographic separation of the lichen extracts.

The experiments with *E. coli* showed the presence in the lichens of substances having vitamin B<sub>12</sub>-like activity in a concentration of from less than 0.005 to 0.035  $\mu\text{g}$  B<sub>12</sub>/g dry weight. This comparatively low activity was found to be due to three different vitamin B<sub>12</sub>-factors having the same  $R_F$ -values as vitamin B<sub>12</sub>,

Table 2. *L. Citrovorum* and *S. faecalis* factors in lichens.

Factors	$R_F$ -values	<i>L. citrovorum</i> activity in samples		<i>S. faecalis</i> activity in samples	
		A	ET	A	ET
Unidentified	0.07			+	+
Pteric acid	0.10				
Unidentified	0.18		++++	+	+++
Pteroyltriglutamic acid	0.22			+	++
Unidentified	0.24	+	+		
Folic acid	0.27			(+)	(+)
Unidentified	0.37			+	+
Formylpteroylglutamic acid	0.45			+++	
Folinic acid	0.55	++	+	++	+
Formylptericoic acid	0.65			+	++
Thymidine	0.70	+	+	+	+

The number of plus signs indicates the intensity of the growth caused by the different factors as observed after chromatographic separation. A = autoclaved, ET = enzyme treated.

vitamin B<sub>12</sub>,<sup>4</sup> and factor C<sup>5,6\*</sup>, respectively. No evidence was found for the presence of factors having the same  $R_F$ -values as pseudovitamin B<sub>12</sub>,<sup>7</sup> vitamin B<sub>12</sub>,<sup>8</sup> vitamin B<sub>12m</sub>,<sup>9</sup> factor A<sup>5</sup>, or factor B<sup>5</sup>. The presence of vitamin B<sub>12</sub> is of interest because to date its only other source has been marine algae<sup>4</sup>.

For *L. citrovorum* four growth factors were found. Two of these have been identified as the natural citrovorum factor<sup>10,11</sup> (folinic acid) ( $R_F$  0.55) and thymidine ( $R_F$  0.70). The other two factors had  $R_F$ -values of 0.18 and 0.24. It seems likely that these factors are identical with two of the citrovorum factors other than folinic acid found in a red marine algae *Furcellaria fastigiata*<sup>12</sup>.

Altogether nine substances stimulating the growth of *S. faecalis* were observed. Folinic acid and thymidine, which stimulate the growth of both *S. faecalis* and *L. citrovorum*, and factors with the same  $R_F$ -values as pteroyltriglutamic acid (Terop-terin, Lederle), N<sub>10</sub>-formylfolic acid and N<sub>10</sub>-formylpteroic acid (Rhizopter-terin) were found. Only traces of folic acid (pteroyltriglutamic acid) could be detected. The other factors observed had approximately the  $R_F$ -values 0.07, 0.18, and 0.37 respectively in this solvent system.

Enzymatic treatment of the extracts did not lead to release of folic acid (pteroyltriglutamic acid) or to folinic acid (N<sub>5</sub>-formyl-tetrahydrofolic acid), but substances with lower  $R_F$ -values. It was also observed that the enzyme complex of the chicken pancreas homogenate did not seem to release folic acid from a solution of crystalline pteroyltriglutamic acid (Terop-terine, Lederle). This is in agreement with previous findings<sup>14,15</sup>. Instead it converted the pteroyltriglutamic acid to a growth factor for *L. citrovorum* which is not folinic acid. The value of the total activity for

\* A factor found in sewage sludge<sup>6</sup> and showing the properties of factor C has been used for comparison.

*S. faecalis* and *L. citrovorum* in Table 1 do not therefore represent the true folic acid or folinic acid content of the lichens.

Most lichens investigated contained substances having an antibiotic effect on *E. coli*, *S. faecalis* and *L. citrovorum*, possibly due to usnic acid.

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